



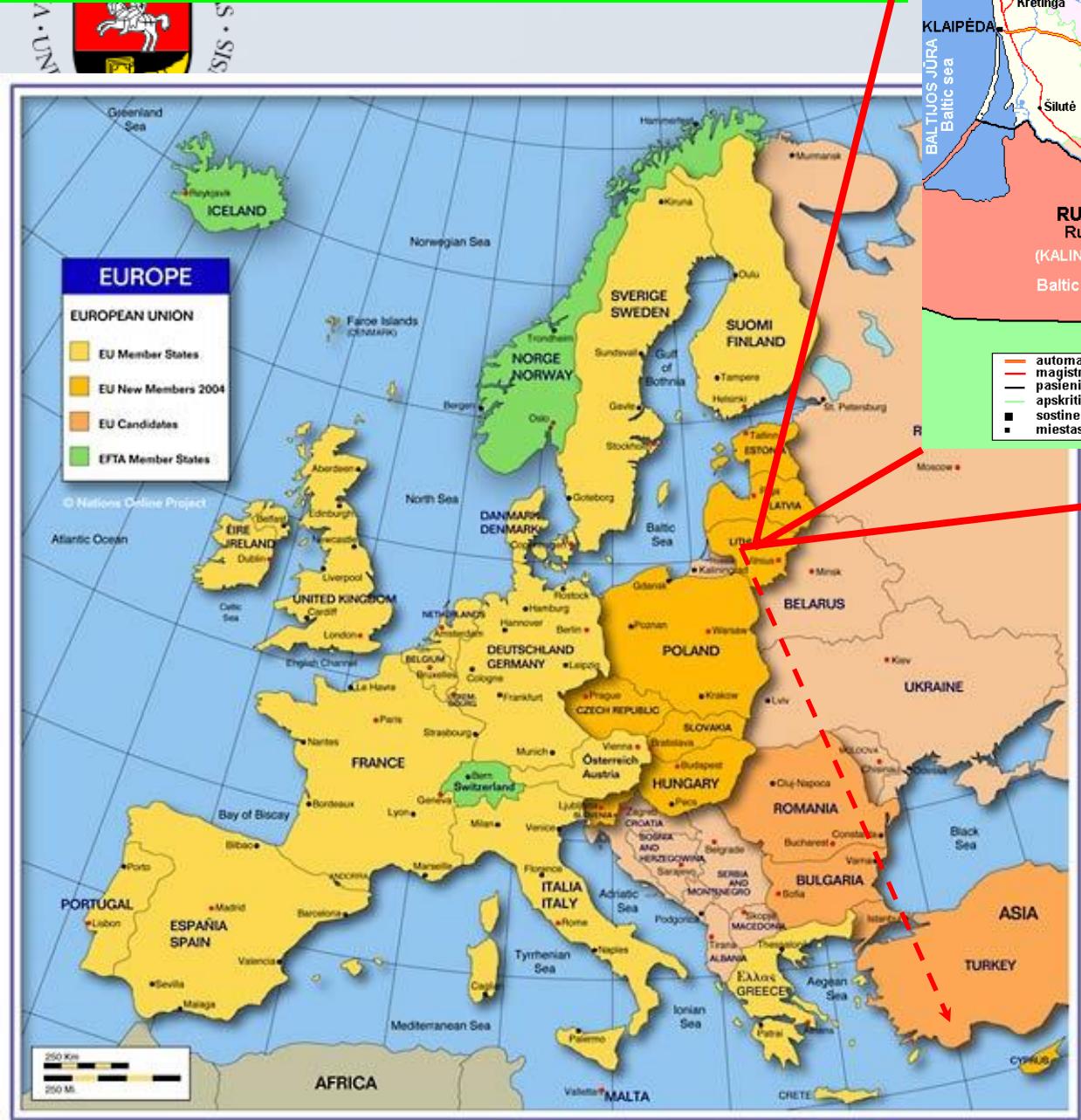
Electron transfer pathways in electrochemical biosensors and in some other biodevices

Prof. dr. habil. Arūnas Ramanavičius

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2. *Laboratory of NanoBioTechnology, Department of Materials Science and Electronics, Institute of Semiconductor Physics, State Scientific Research Institute Centre for Physical Sciences and Technology. Gostauto 9, LT-01108 Vilnius, Lithuania.*



Lithuania (Lietuva)



Capital: Vilnius

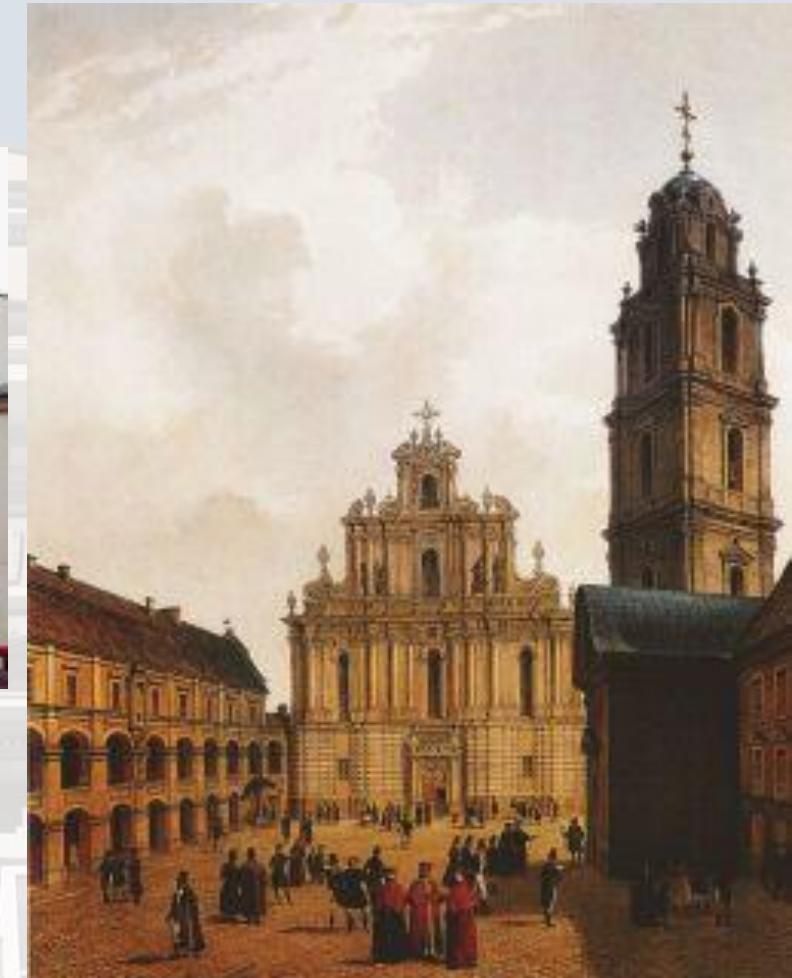
3.0 Mln. People.





Vilnius University

Was founded in 1579

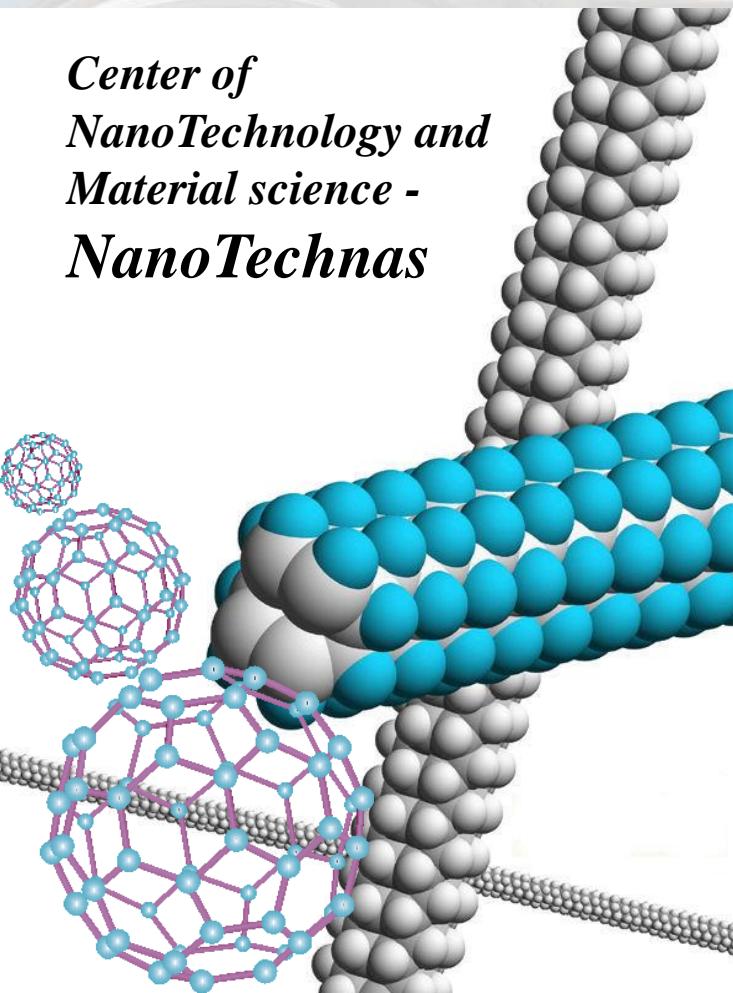




Center of NanoTechnology and Materials science - NanoTechnas

Faculty of Chemistry, Vilnius University

*Center of
NanoTechnology and
Material science –
NanoTechnas*



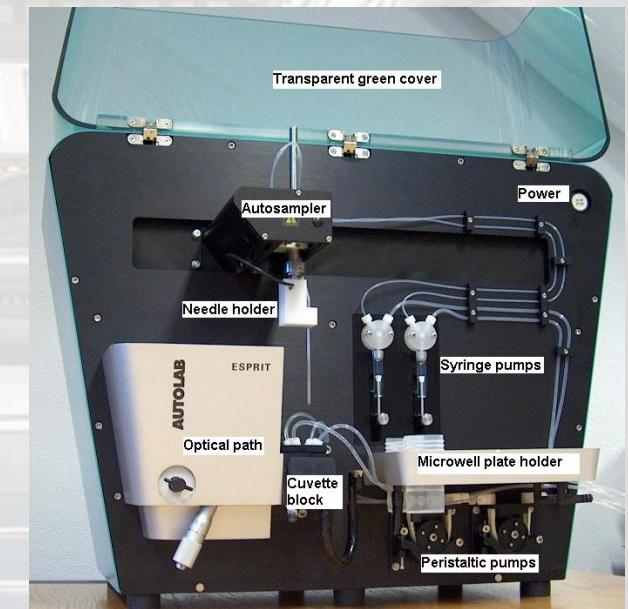
**Center of NanoTechnology
and Material science –
NanoTechnas**
established within 2005-2008



Some Research Directions in NanoTechnas

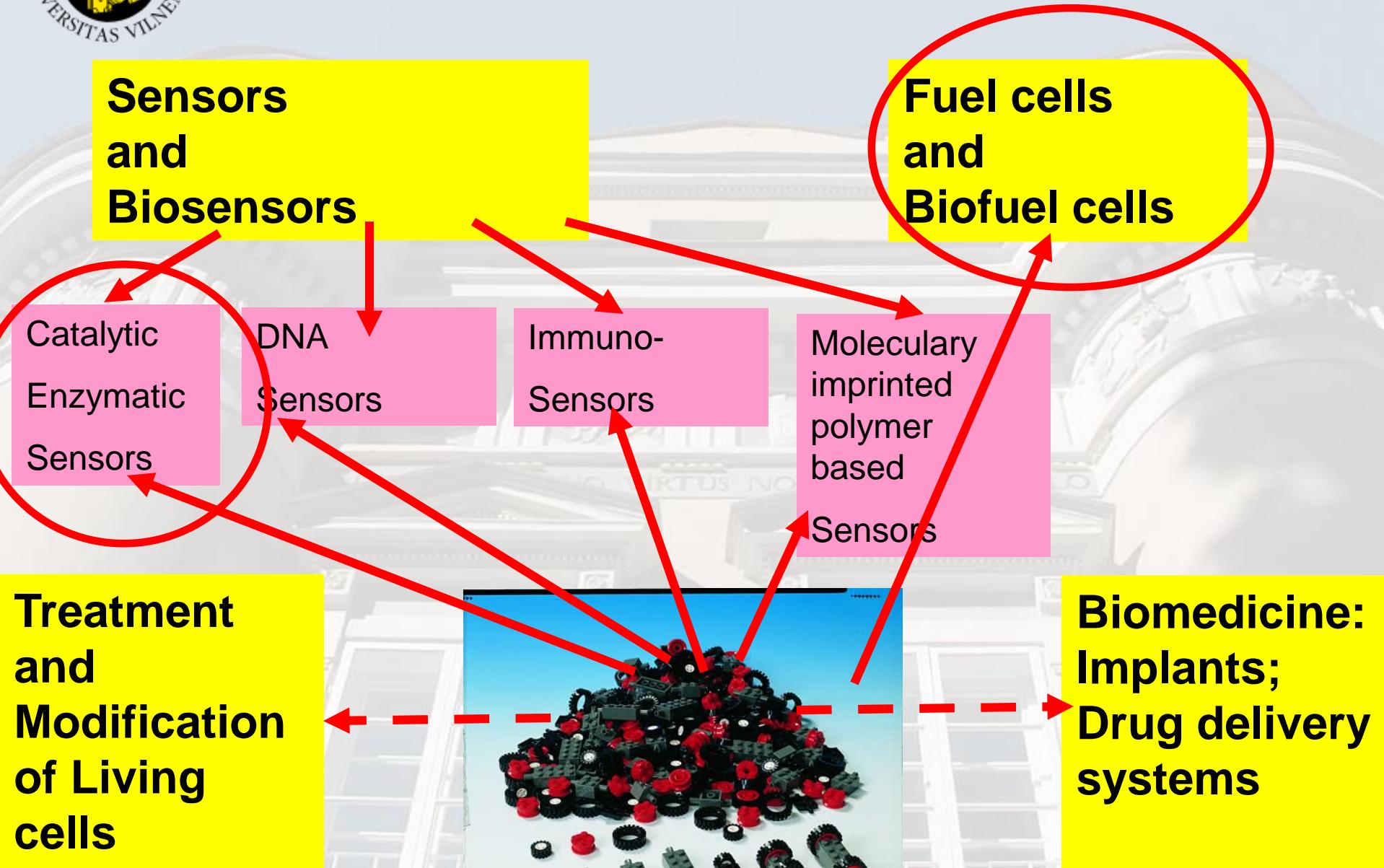


- Bio – AFM (Veeco)
- SPR device (ECO-Chemie);
- QCM+EQCM (MacTex);
- Potentiostat/Galvanostat (ECO-Chemie);





Some Research Directions in NanoTechnas



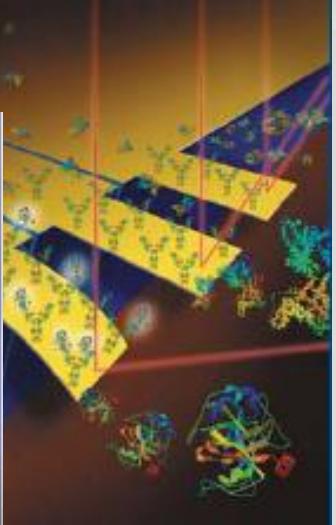
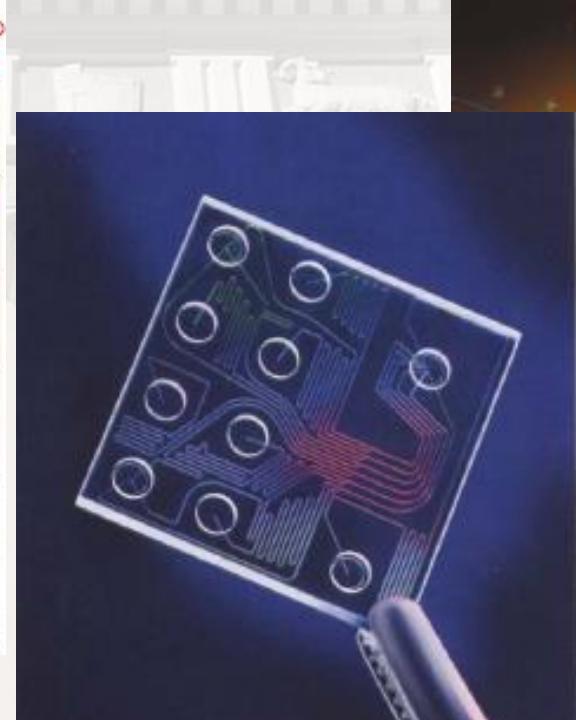


Introduction // Sensors // Lab on the chip

Do we need sensors?

Why do we need sensors?

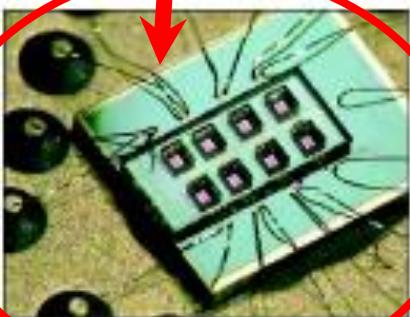
Sensors // What they are??



Introduction // Some Challenges in Sensorics and Biosensorics // Biofuel Cells in Biosensors

Need:
Miniaturization
& Integration

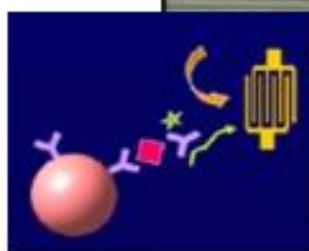
*Signal Processing
& Transmission*



*Sensing
Element
Array*



7.5 μm (width and space)
Reference electrode
IDA
Auxiliary electrode



$\sim\text{nm}$

$\sim\mu\text{m}$



$\sim\text{cm}$



$\sim\text{mm}$



An Integrated Sensor

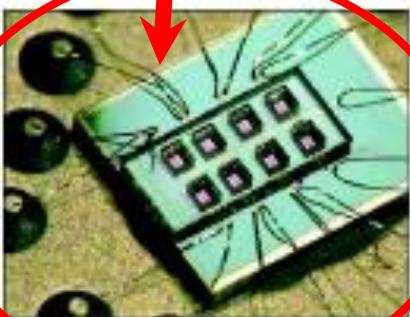
- Sample Delivery
- Sensing Elements
- Signal Processing & Transmission



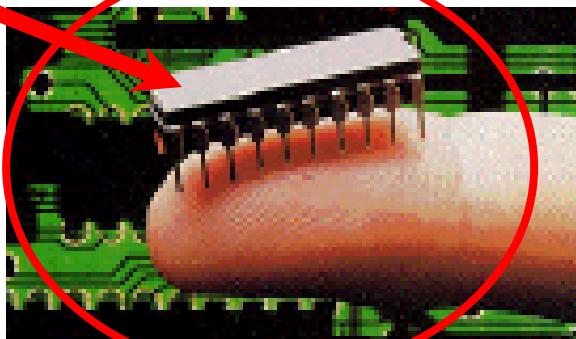
Application of Biofuel Cells in Biosensorics

Need:
Miniaturization
& Integration

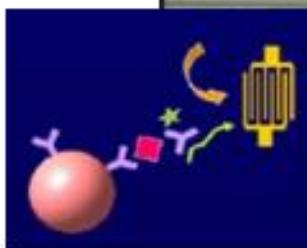
Signal Processing
& Transmission



Sensing
Element
Array



Reference
electrode
IDA
Auxiliary
electrode



~nm

~ μ m



~cm

~mm

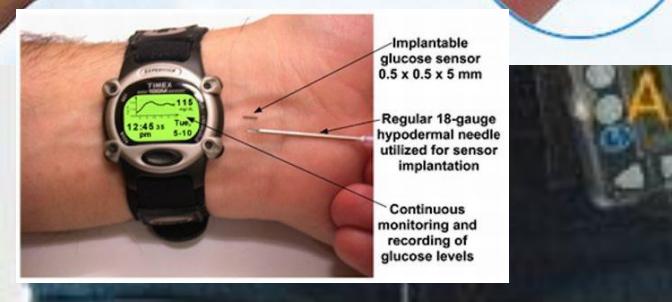
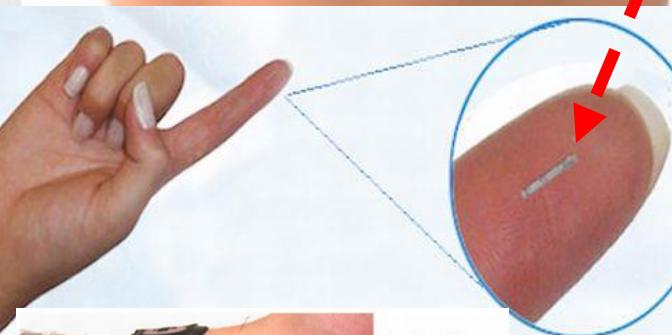
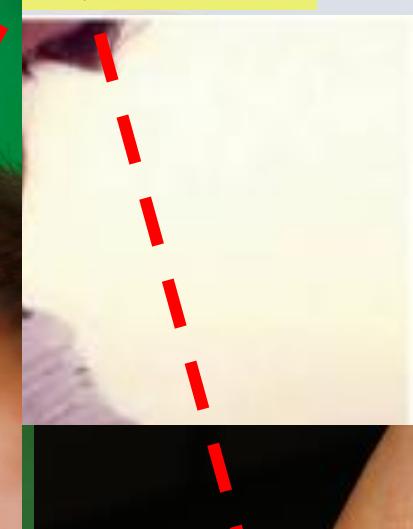
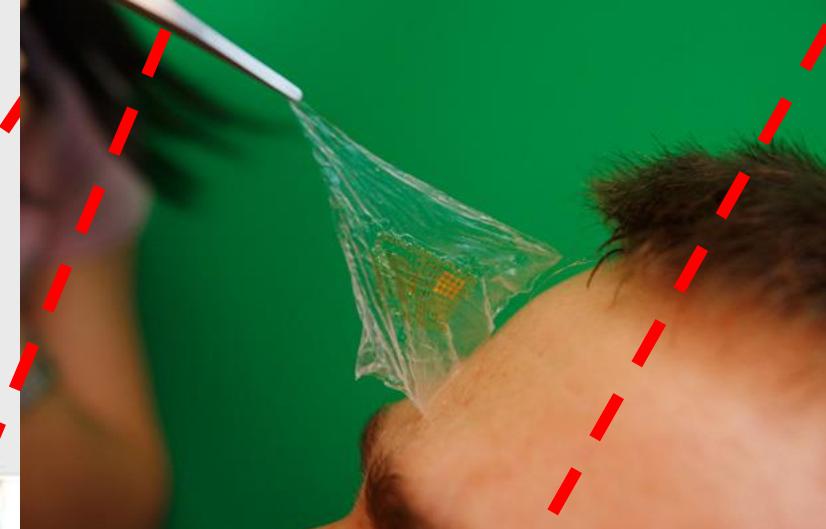


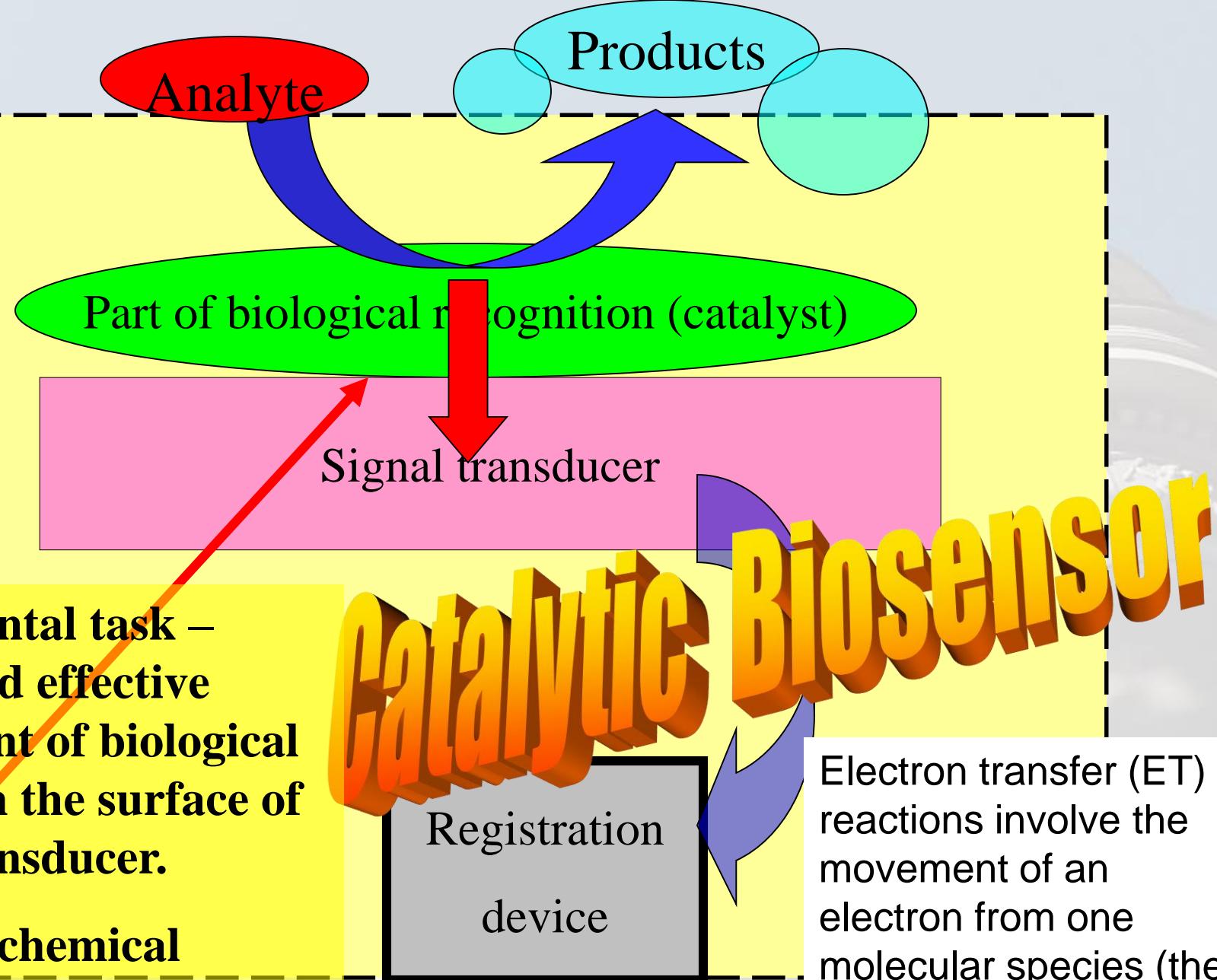
An Integrated Sensor

- Sample Delivery
- Sensing Elements
- Signal Processing & Transmission



Sensors Reality and Dreams





Electron transfer (ET) reactions involve the movement of an electron from one molecular species (the donor) to another (the acceptor)



Enzyme catalyzed reactions

- In addition to many other types of catalysis biochemical catalysis also has number of advantages, e.g.:
- Enzymes are “green catalysts” that are produced by all living cells and/or microorganisms. **Life is impossible without enzymes...**
- COST Action TD1102

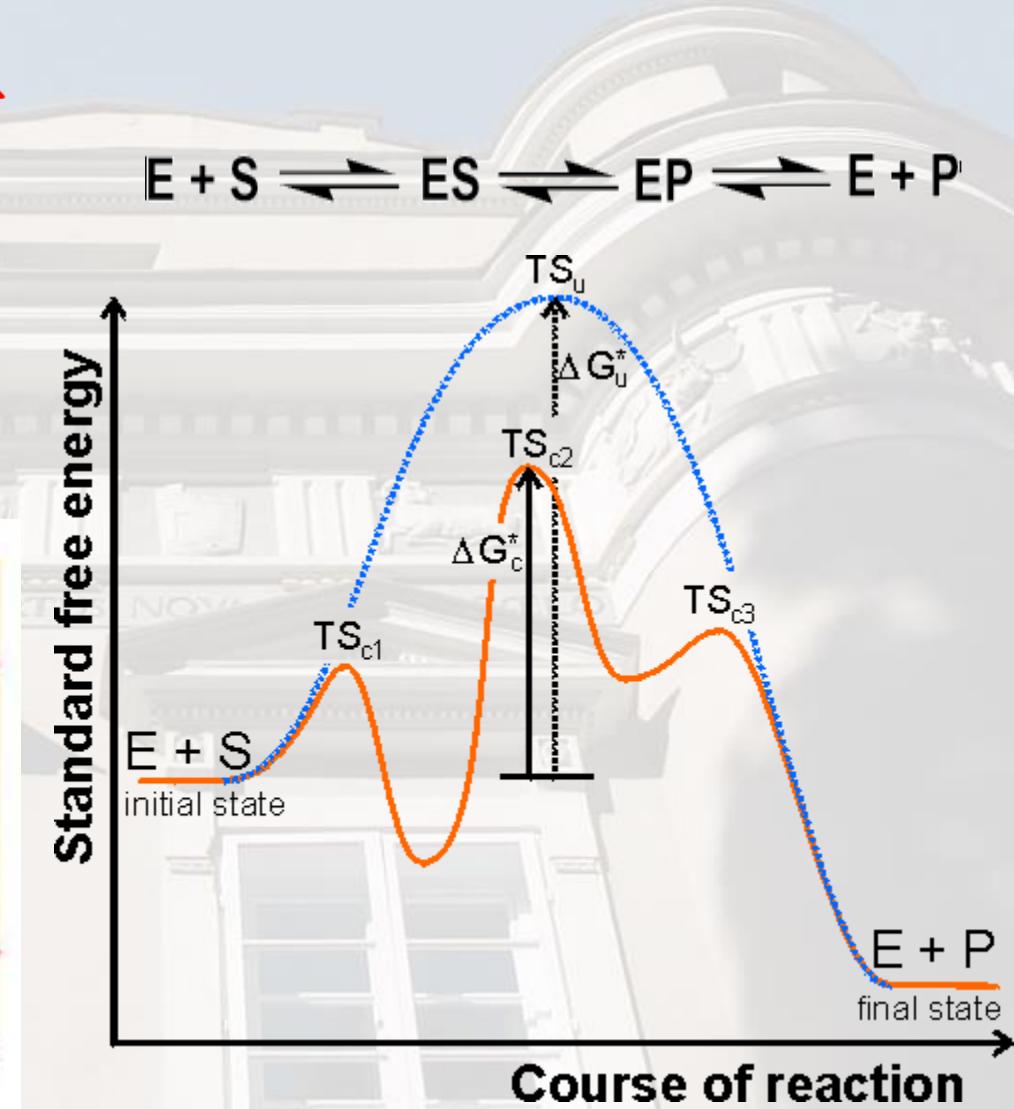
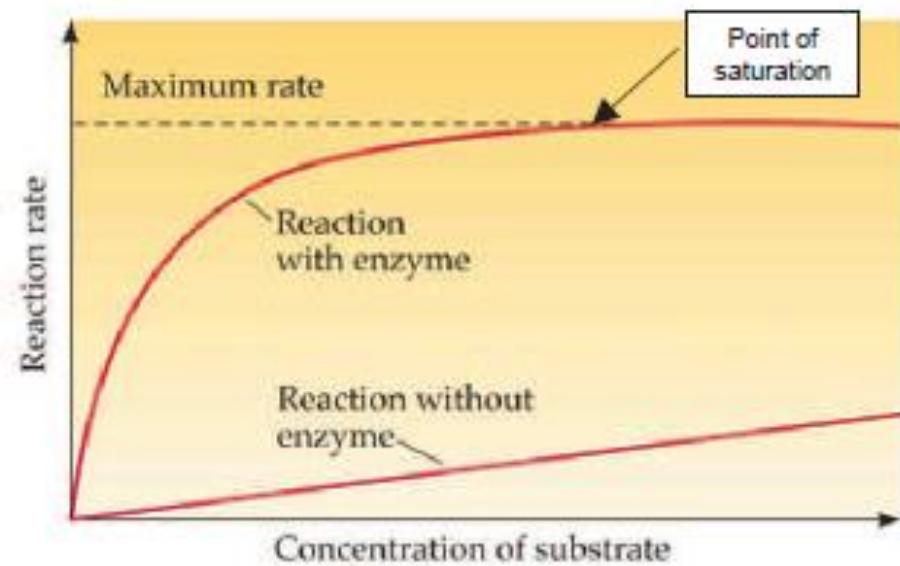
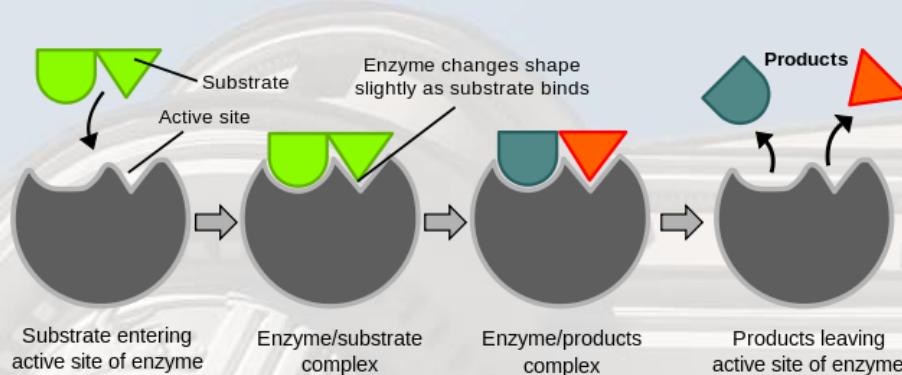


Enzymes are biological catalysts

- There are known about 40,000 different enzymes alone in human cells, each controlling a different chemical reaction. They increase the rate of reactions by a factor of between 10 to 10¹² times, allowing the chemical reactions that make life possible to take place at normal temperatures. They were discovered in fermenting yeast in 1900 by Buchner, and the name enzyme means "in yeast". As well as catalysing all the metabolic reactions of cells (such as respiration, photosynthesis and digestion), they may also act as motors, membrane pumps and receptors.



Enzymatic catalysis

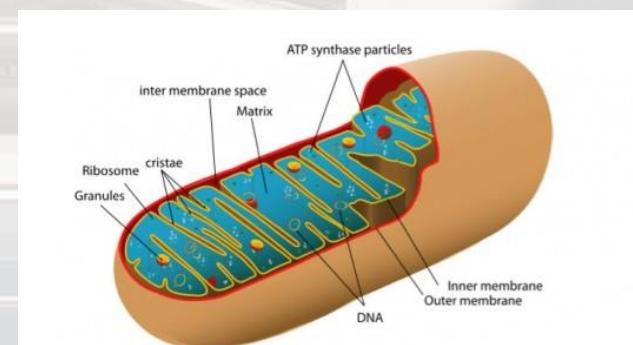
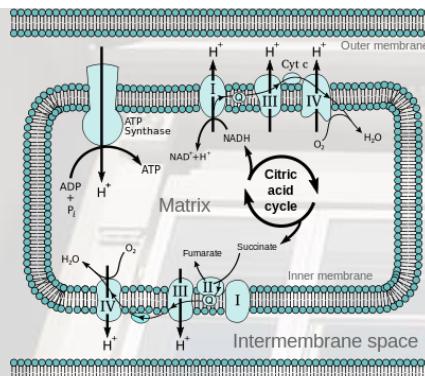
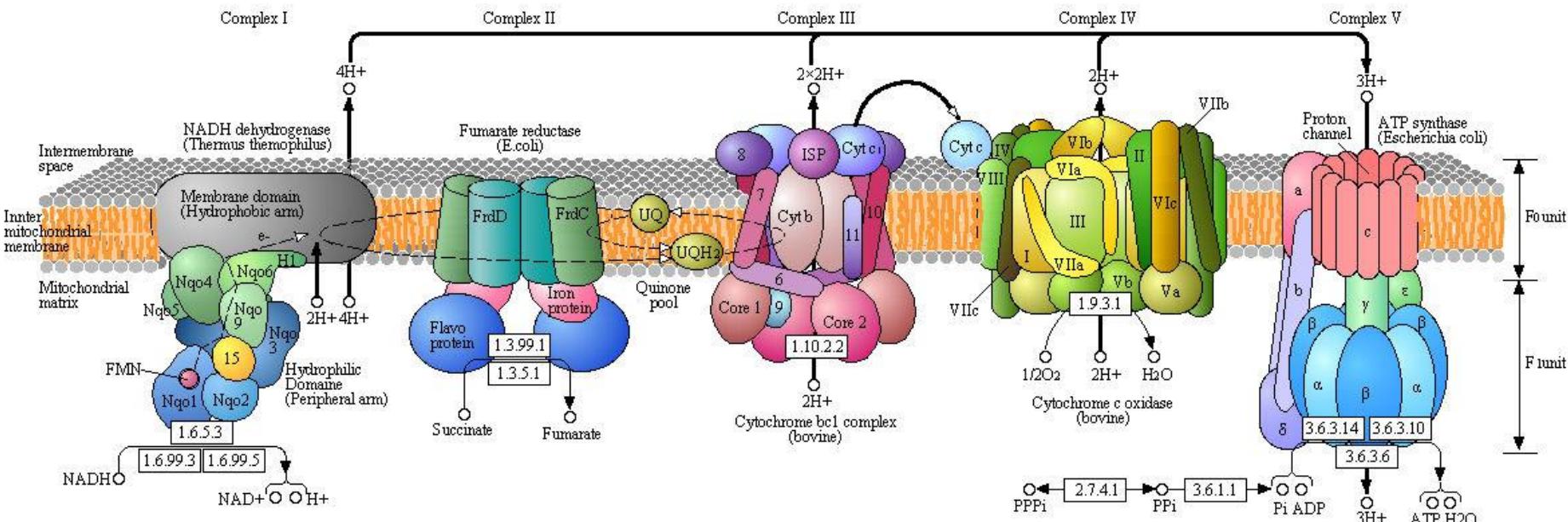


Classes of Enzymes

<u>Class</u>	<u>Chemical Reaction Catalyzed</u>	<u>Sample Enzymes</u>
Oxidoreductase	Oxidation-reduction in which oxygen and hydrogen are gained or lost	Cytochrome oxidase, lactate dehydrogenase
Transferase	Transfer of functional groups, such as an amino group, acetyl group, or phosphate group	Acetate kinase, alanine deaminase
Hydrolase	Hydrolysis (addition of water)	Lipase, sucrase
Lyase	Removal of groups of atoms without hydrolysis	Oxalate decarboxylase, isocitrate lyase
Isomerase	Rearrangement of atoms within a molecule	Glucose-phosphate isomerase, alanine racemase
Ligase	Joining of two molecules (using energy usually derived from the breakdown of ATP)	Acetyl-CoA synthetase, DNA ligase



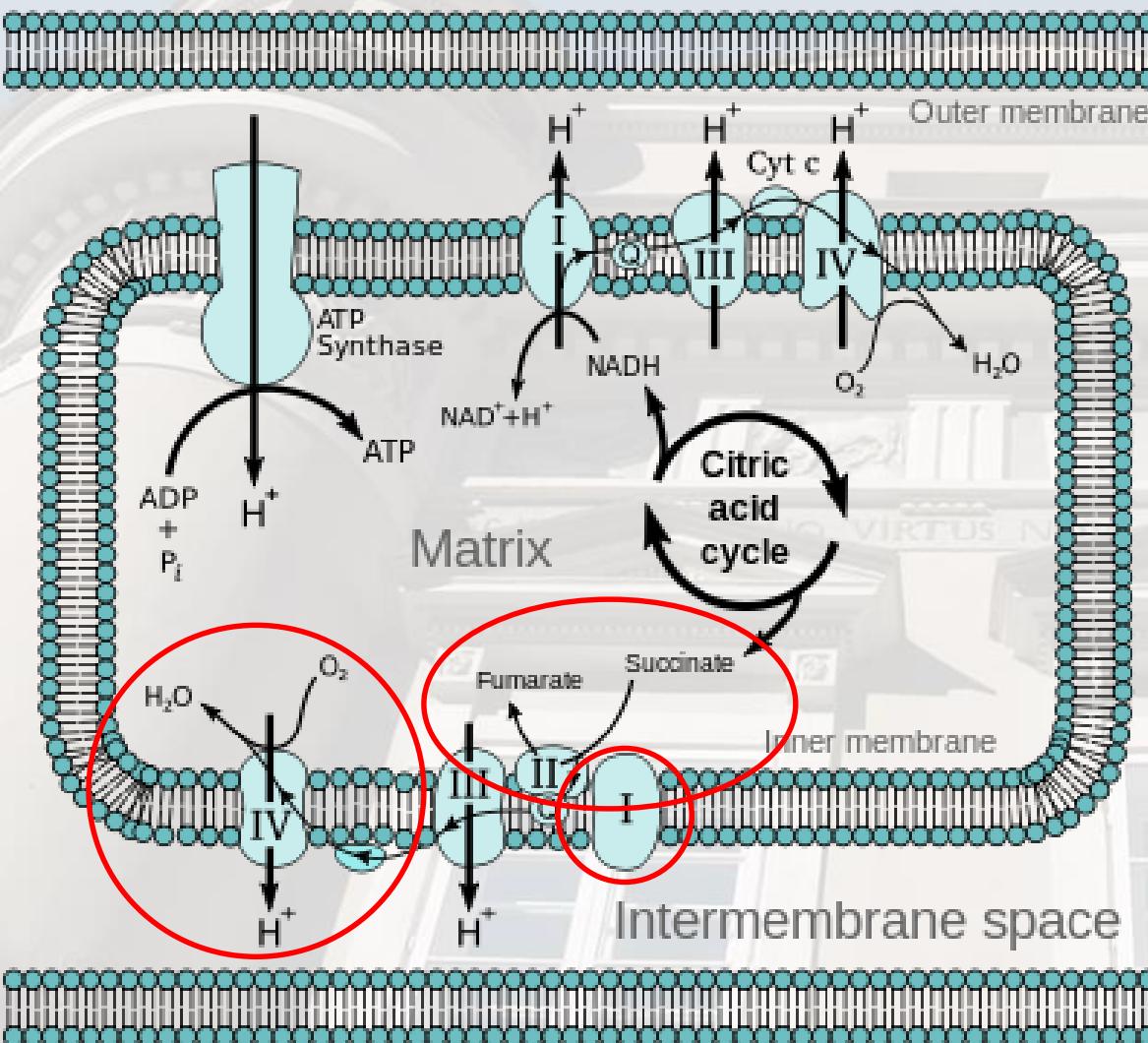
Some Examples of Enzymatic Redox Reactions





Some Examples of Enzymatic Redox Reactions

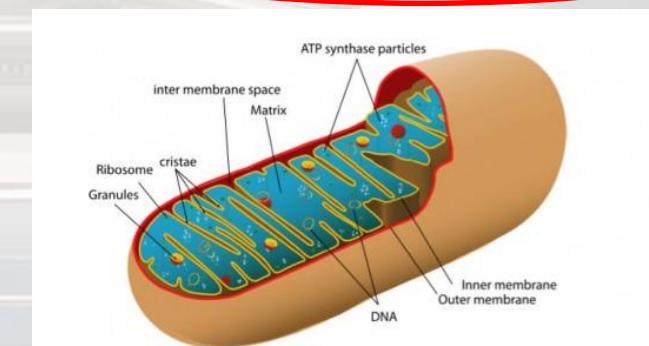
(Oxidative phosphorylation)



Complex I or NADH-Q oxidoreductase.

Complex II: Succinate-Q oxidoreductase.

Complex IV: cytochrome c oxidase.

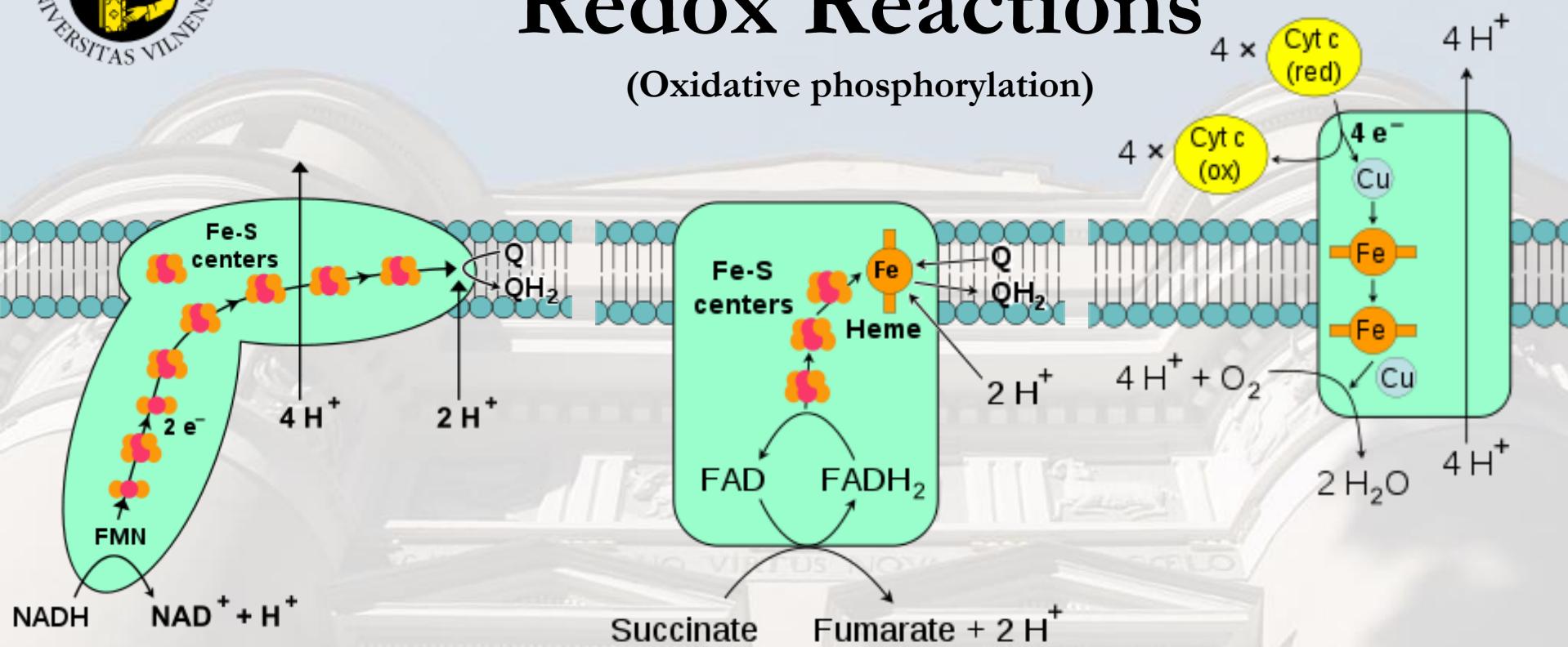


http://en.wikipedia.org/wiki/Oxidative_phosphorylation

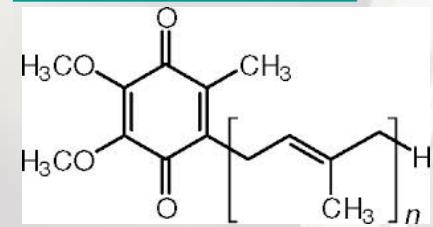


Some Examples of Enzymatic Redox Reactions

(Oxidative phosphorylation)

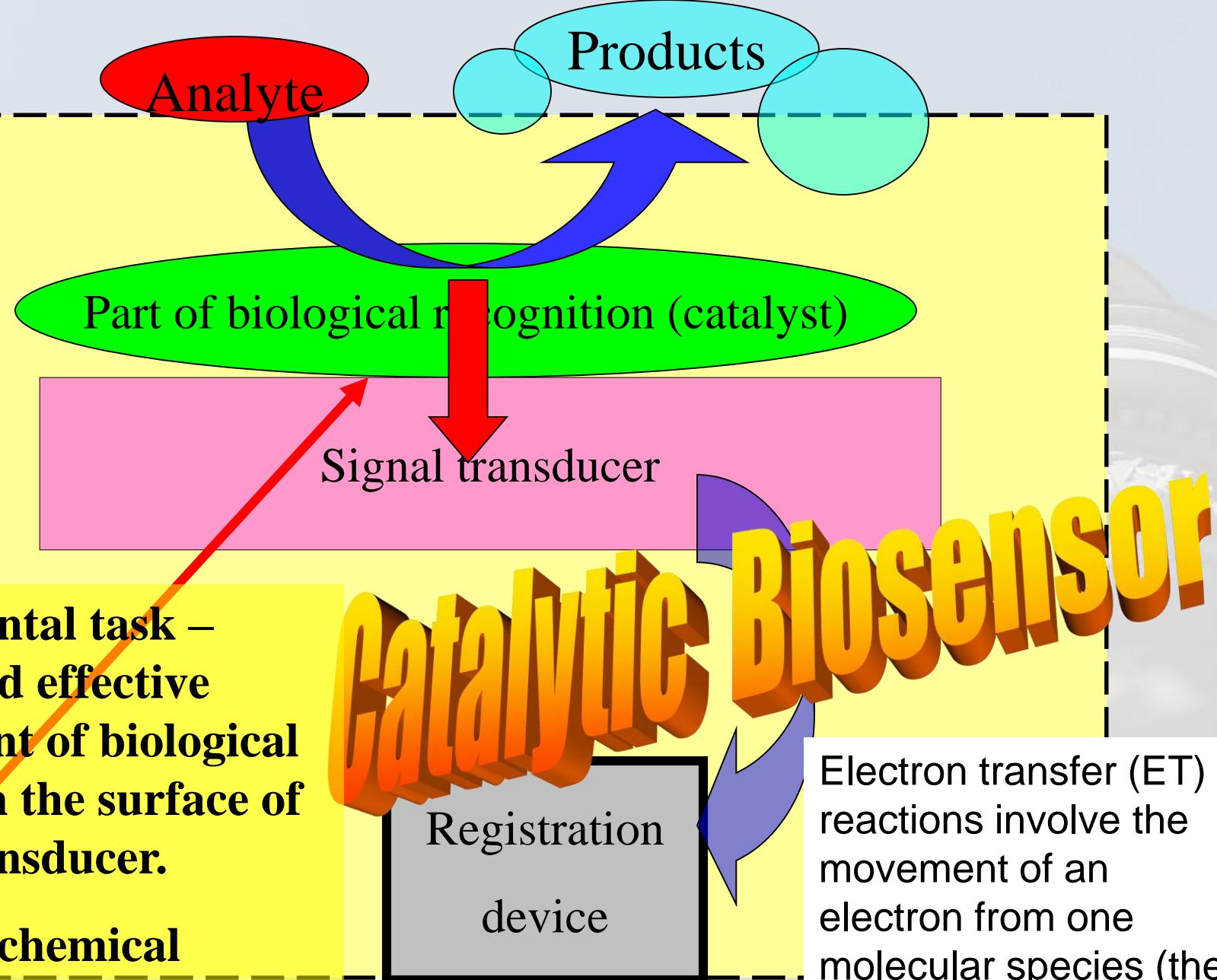


Complex I or NADH-Q oxidoreductase.

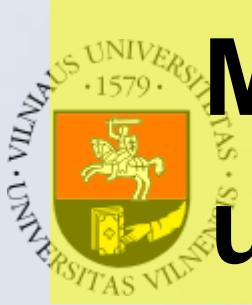


Complex II:
Succinate-Q oxidoreductase.

Complex IV:
cytochrome c oxidase.

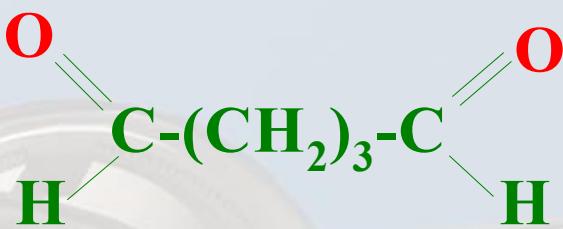


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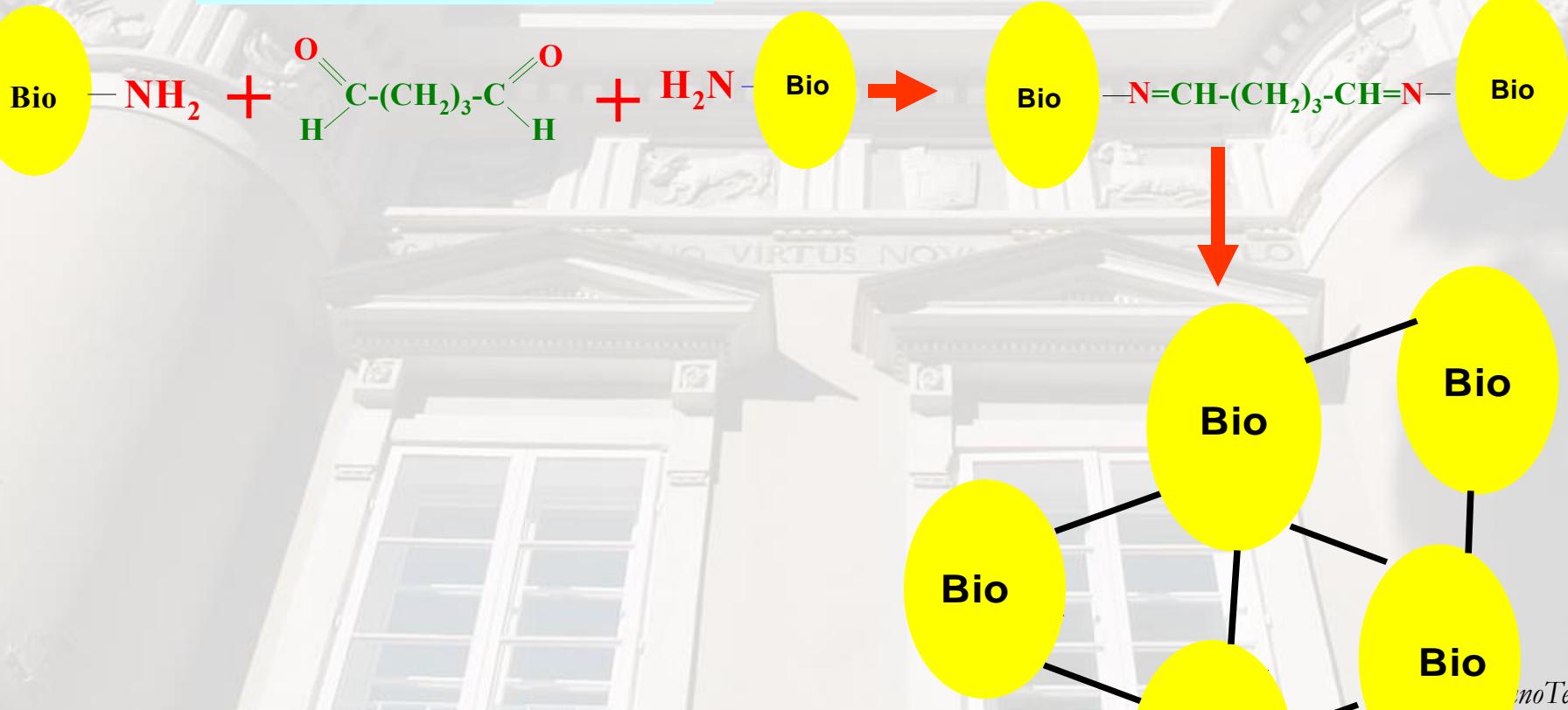


Major immobilization methods used for design of biosensors

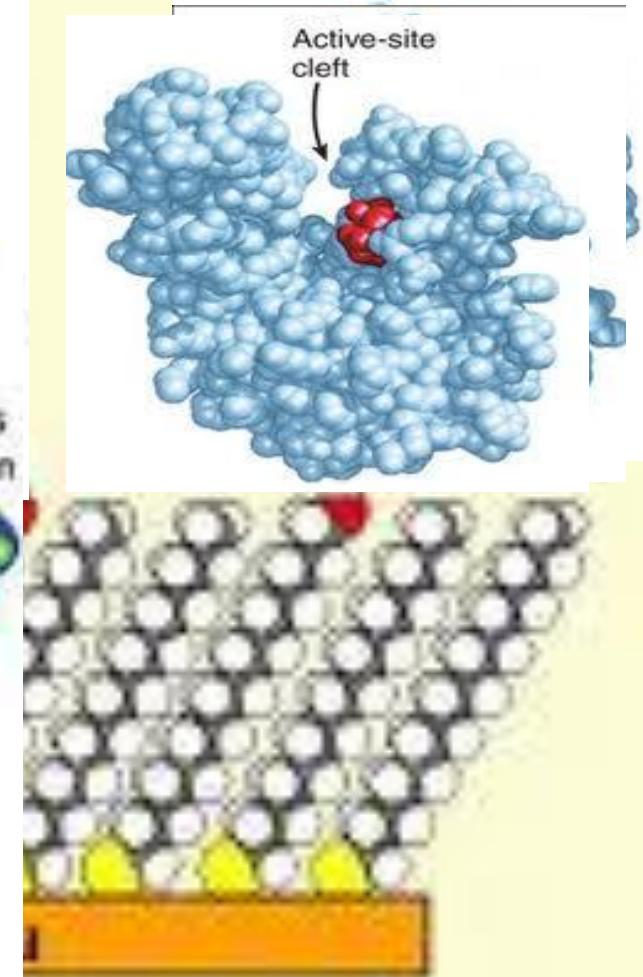
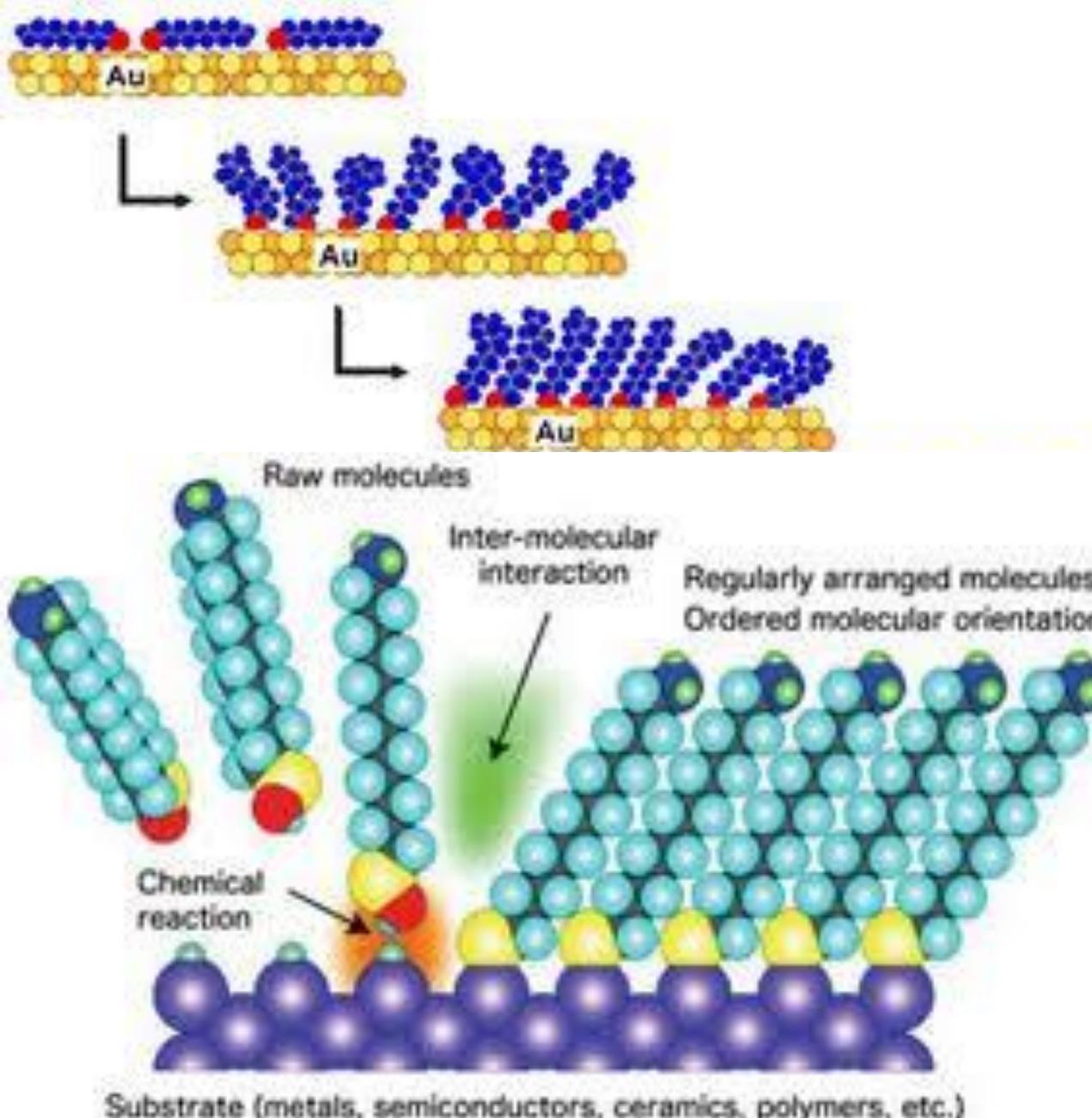
- Adsorption;
- Covalent attachment;
- Cross-linking with chemical agents;
- Application of SAM's followed by covalent attachment;
- Entrapment within polymers.



**Pentandiol
(Glutaraldehyde)**



Immobilization methods cross-linking

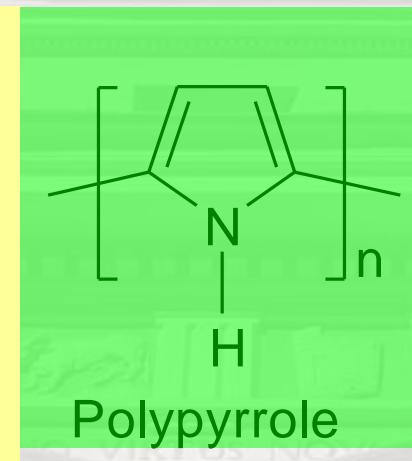


Substrate (metals, semiconductors, ceramics, polymers, etc.)



Entrapment within polymers / Chemical structures of common conducting polymers / Selection principles

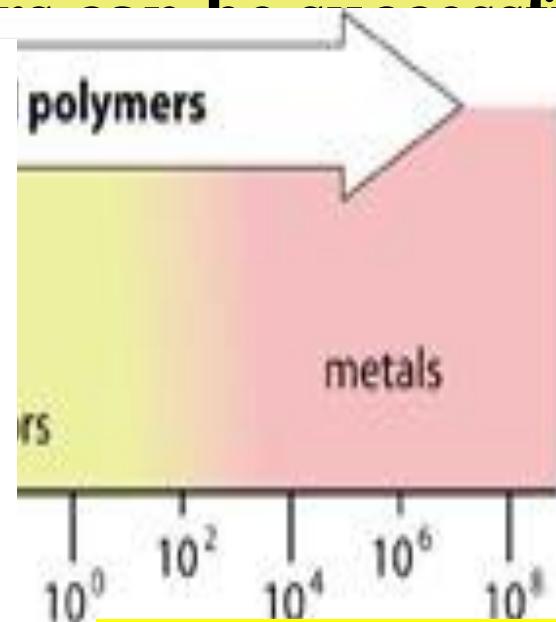
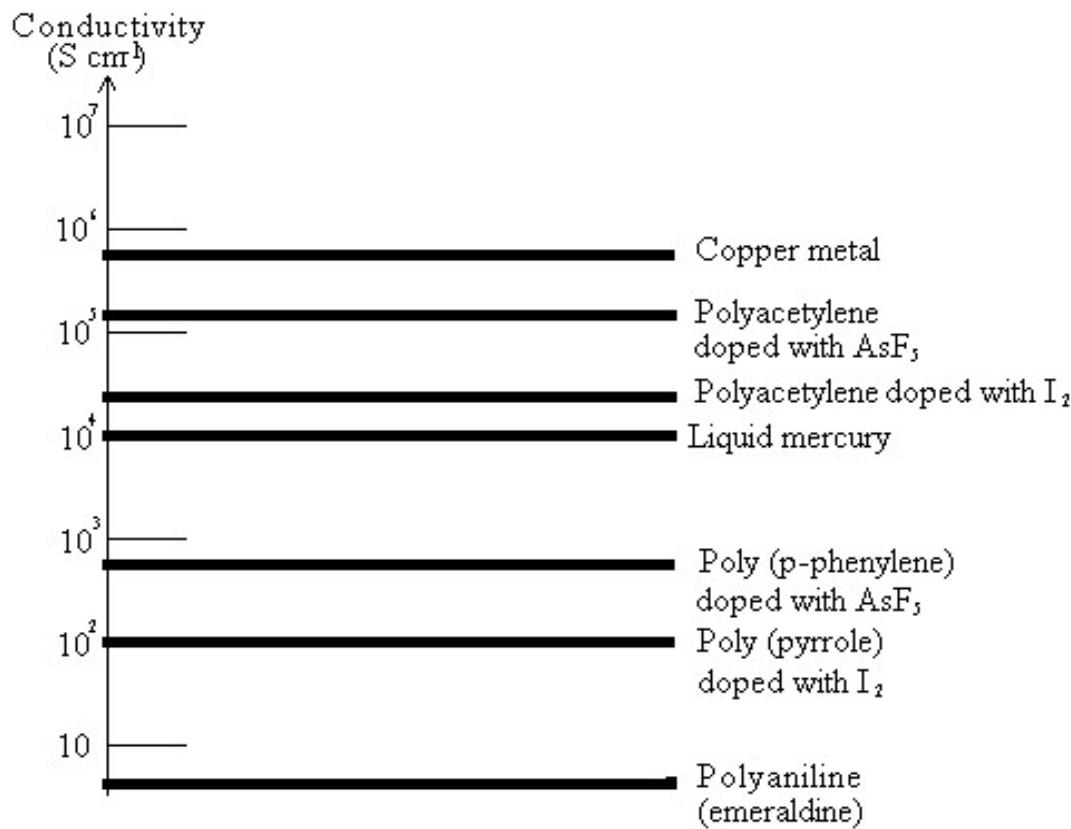
Can be electrochemically synthesized on the conducting surfaces.



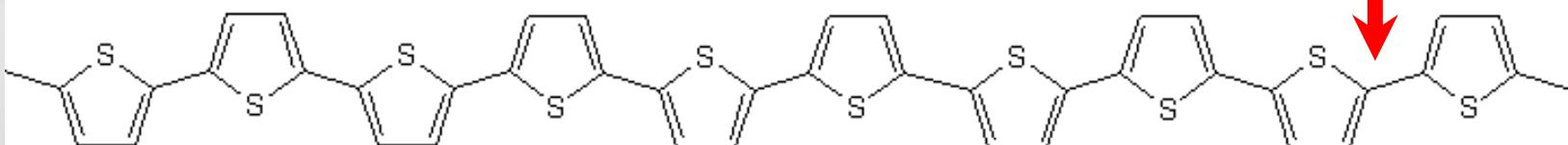
The layer is stable and elastic

Can be doped by biological objects e.g.: proteins.

Can be synthesized at neutral pH.
(It is especially important during immobilization of biological objects)

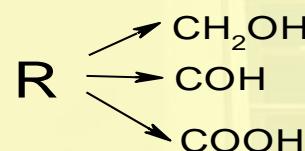
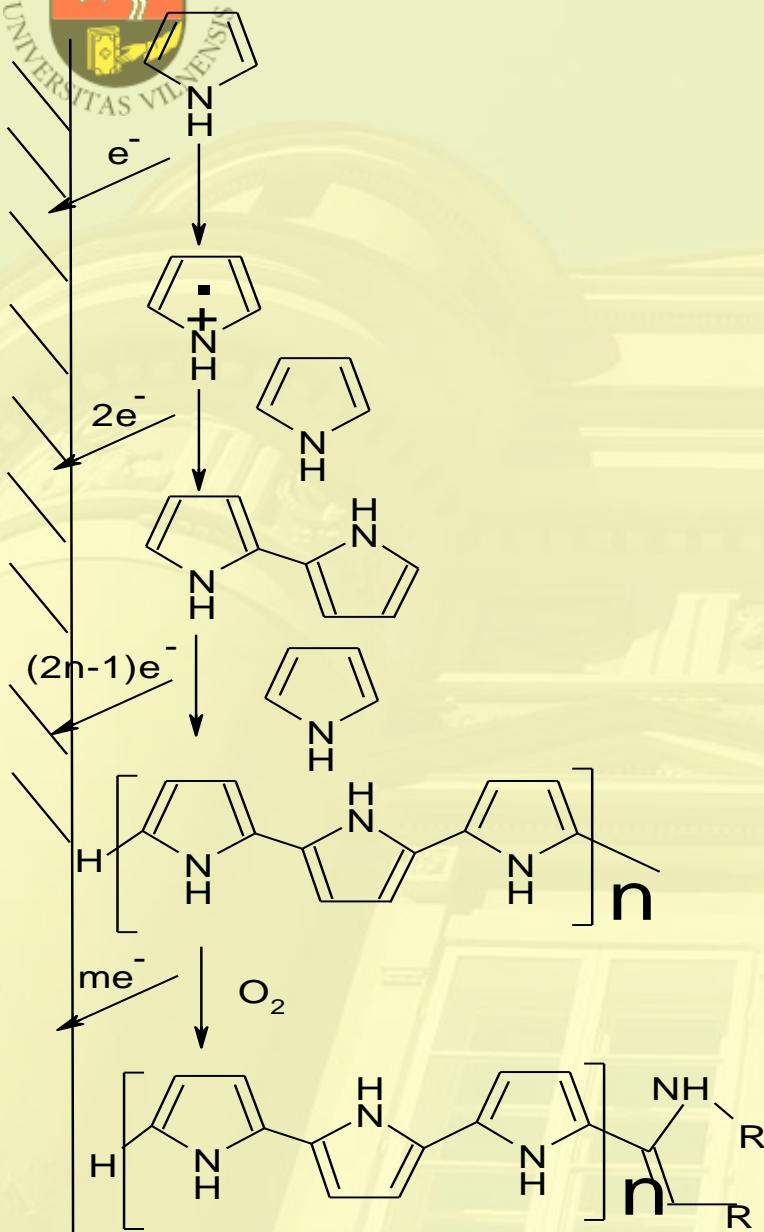


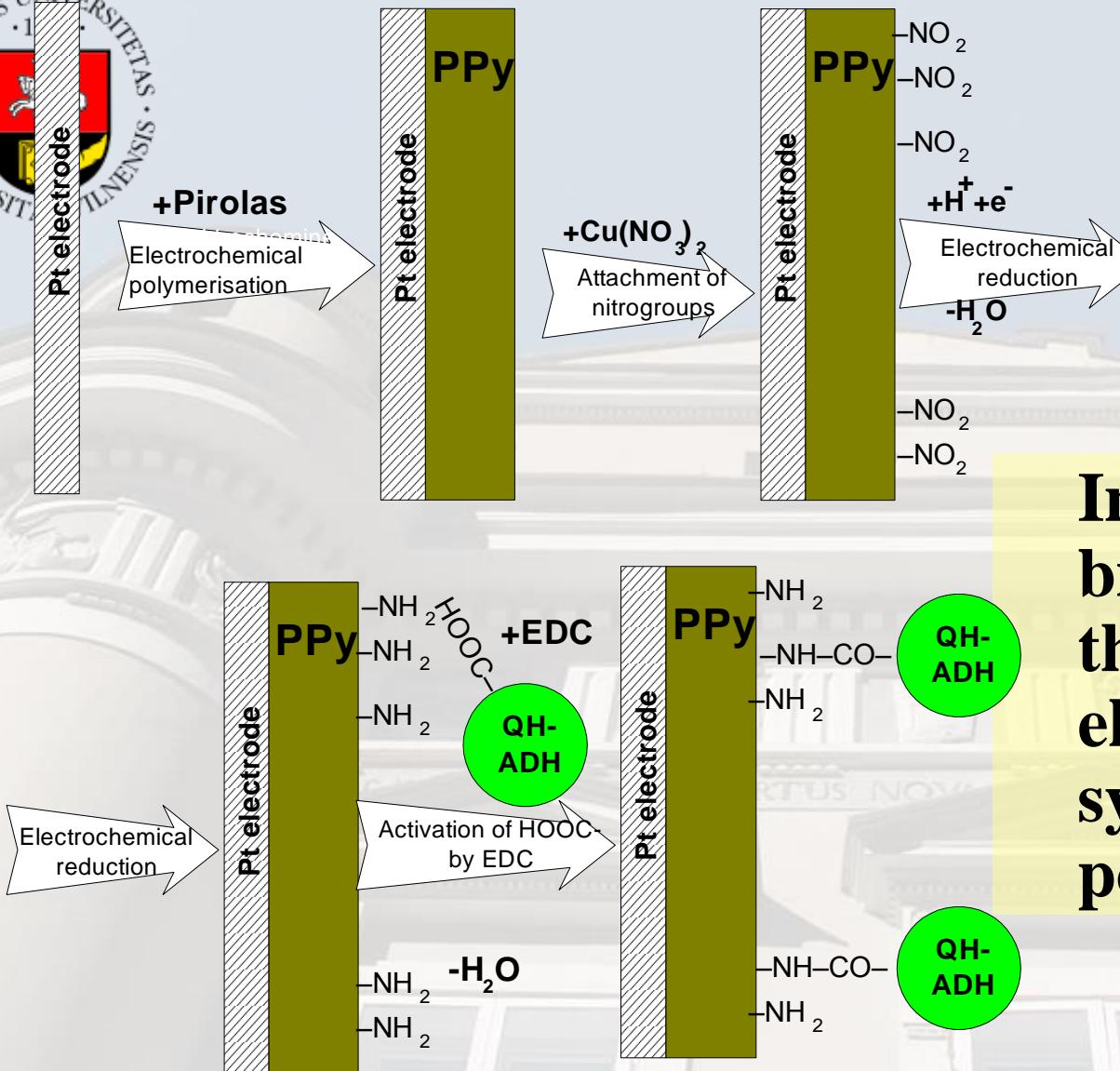
Charge transfer via conjugated polymeric backbone





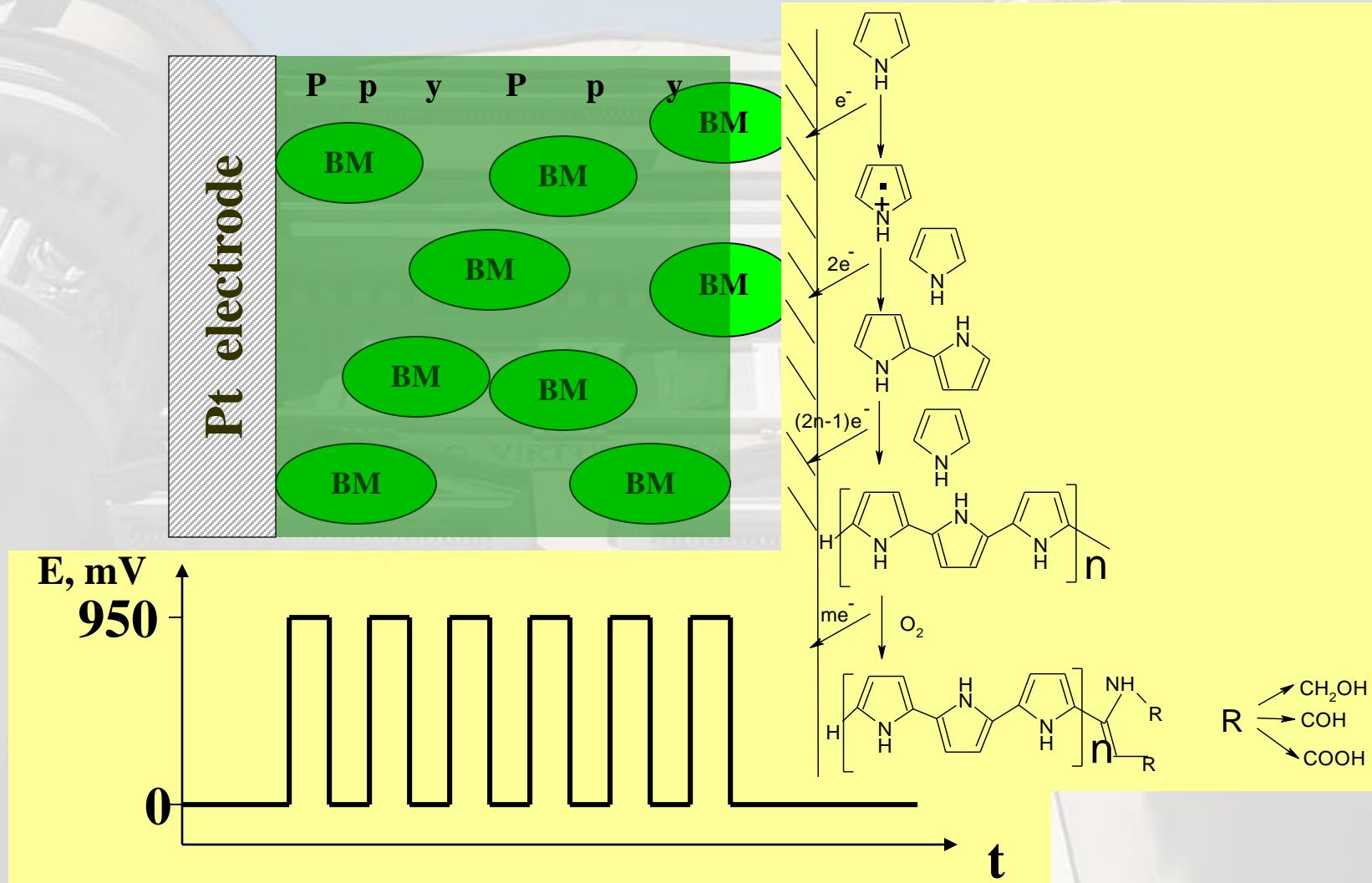
The Electrochemical synthesis of conducting polymers (e.g. polypyrrole)



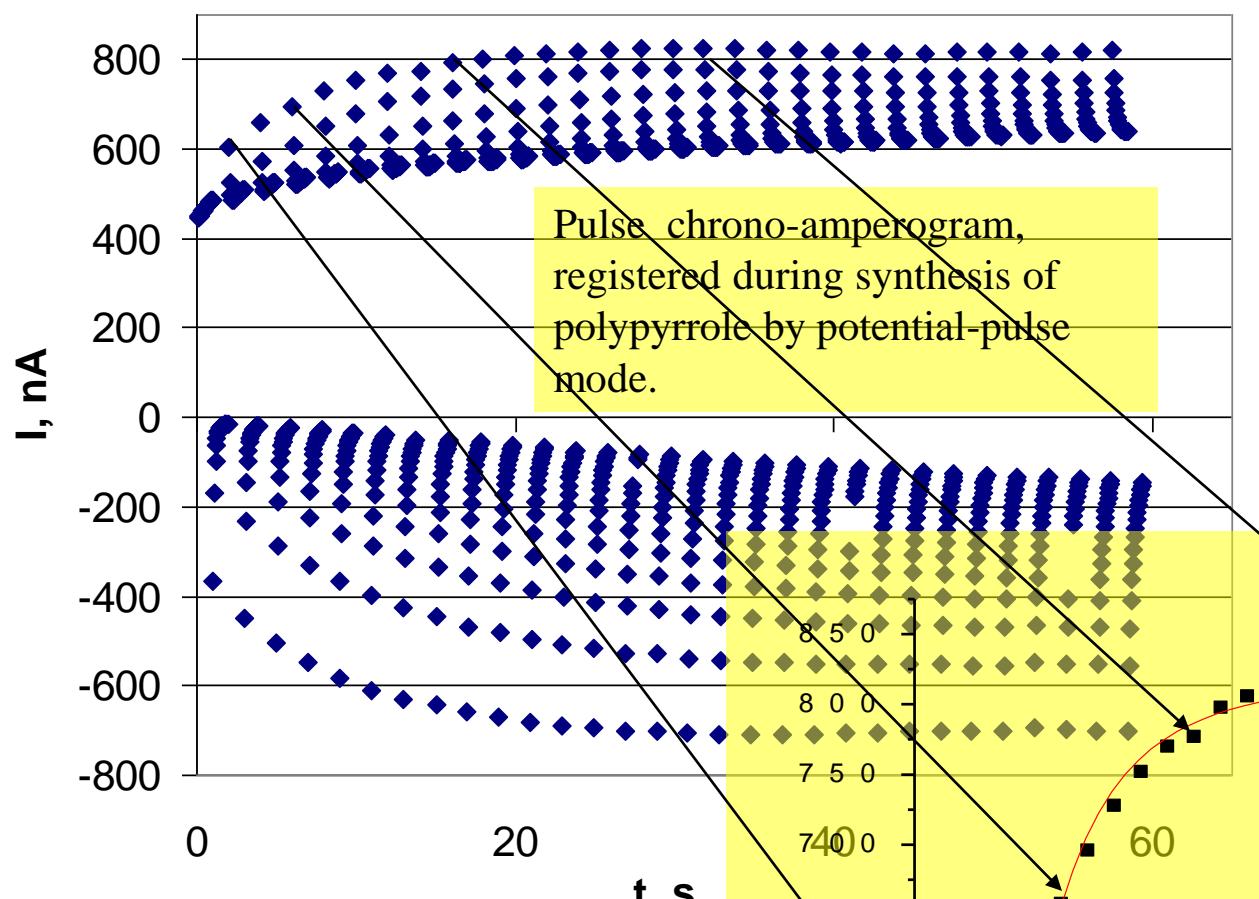


Immobilization of bio-molecules on the surface of electrochemically synthesized polypyrrole

Immobilization of bio-molecules within electrochemically formed polypyrrole (Entrapment of bio-molecules within Ppy)

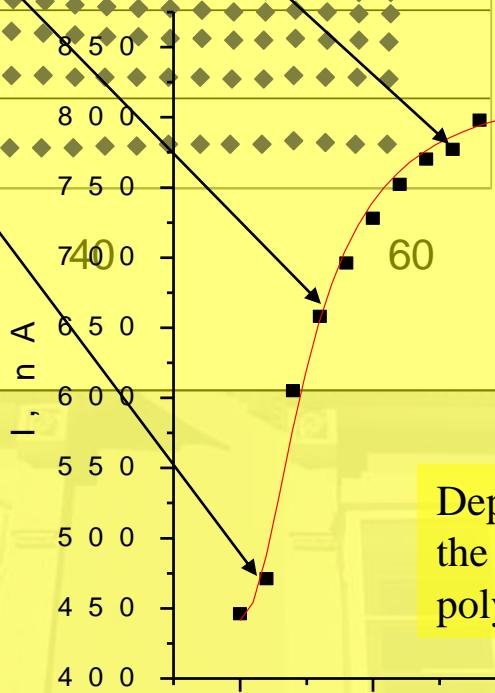


Control of Ppy film growth



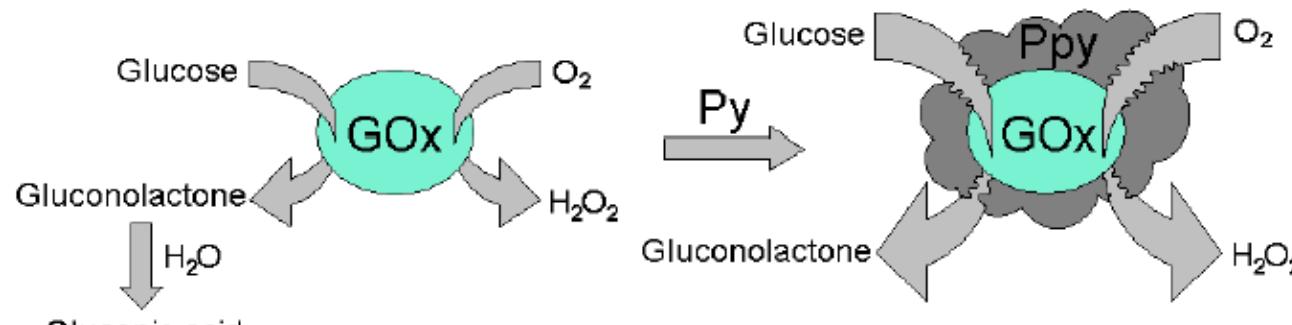
$$y = \frac{Z-K}{1+(L*x)^D} + K;$$

Where:
 $Z=441.0 \pm 8.1$;
 $K=825.0 \pm 2.45$;
 $L=0.3751 \pm 0.0112$;
 $D=2$. (1.95 ± 0.03)

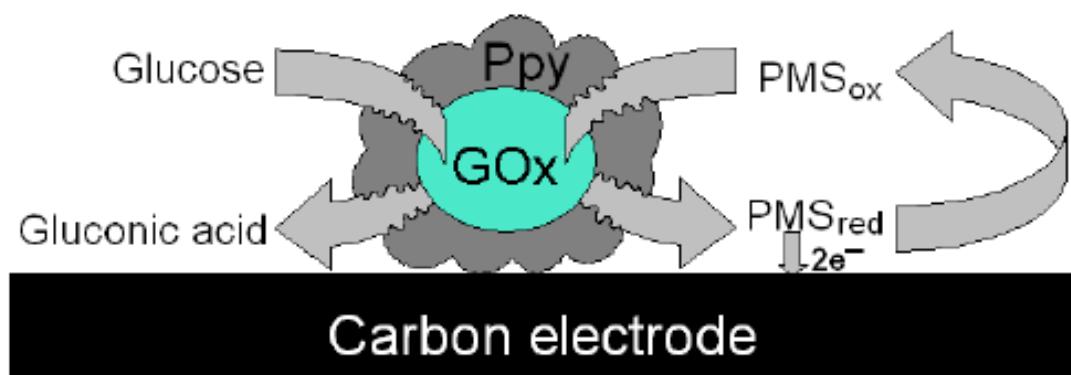


Pulse number

Polypyrrole Coated Glucose Oxidase Nanoparticles

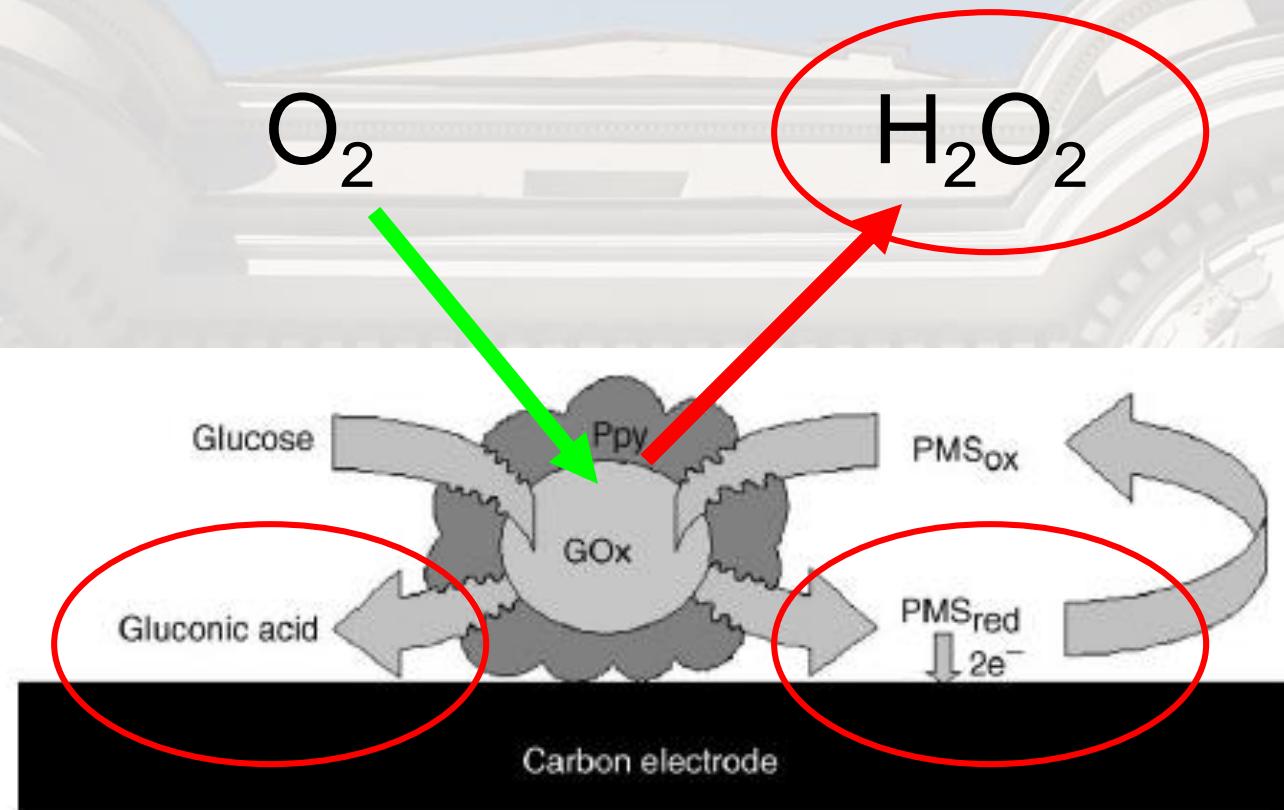


Schematic representation of glucose oxidase coating by polypyrrole initiated by catalytic action of this enzyme.



Schematic representation of application of polypyrrole coated glucose oxidase nanoparticles in PMS mediated biosensor design.

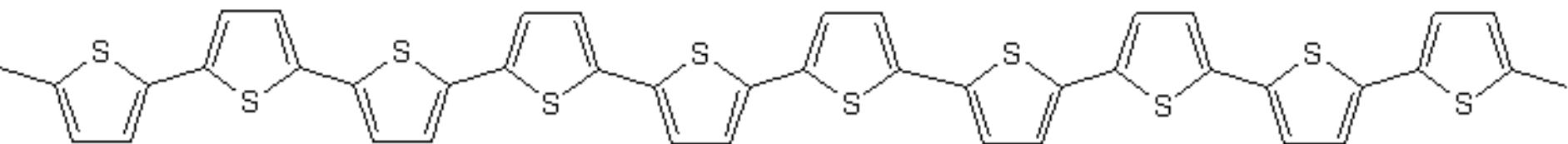
Activity of entrapped enzymes





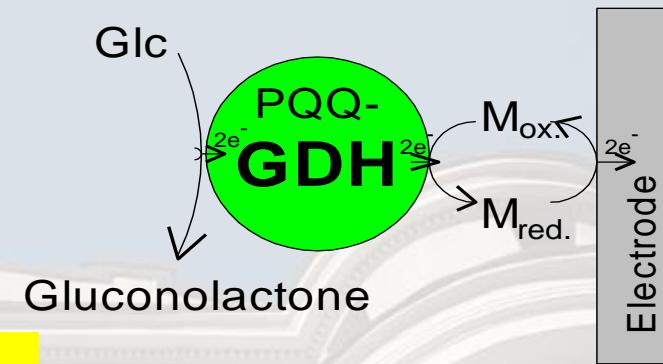
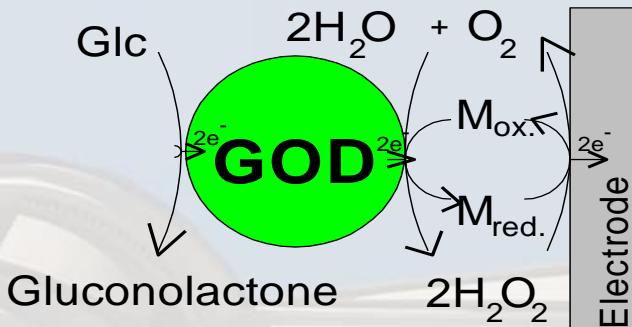
Major immobilization methods used for design of biosensors

- Adsorption;
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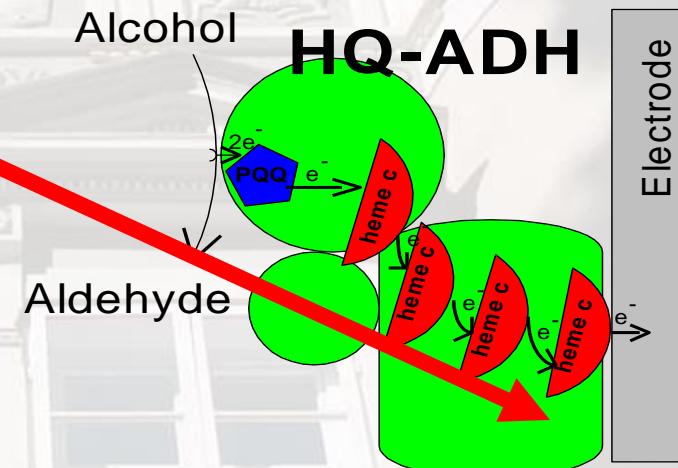
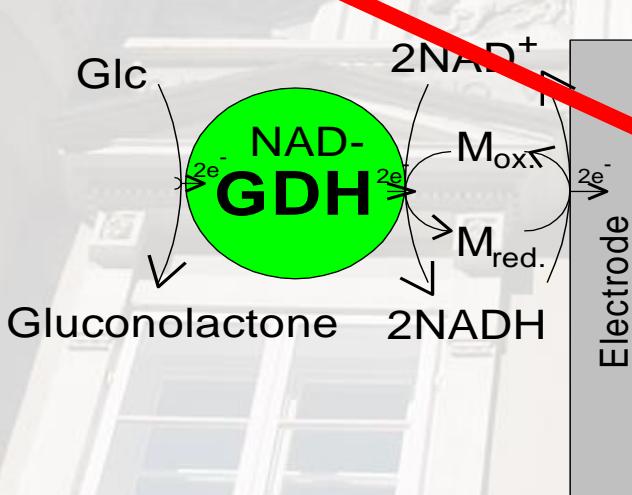
Conducting polymers might be very useful: (i) for modification, (ii) and for protection of biosensor surfaces. (2nd lecture)

Enzymes in design of catalytic biosensors

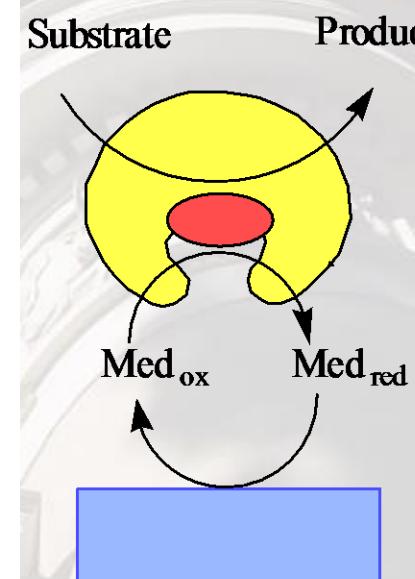
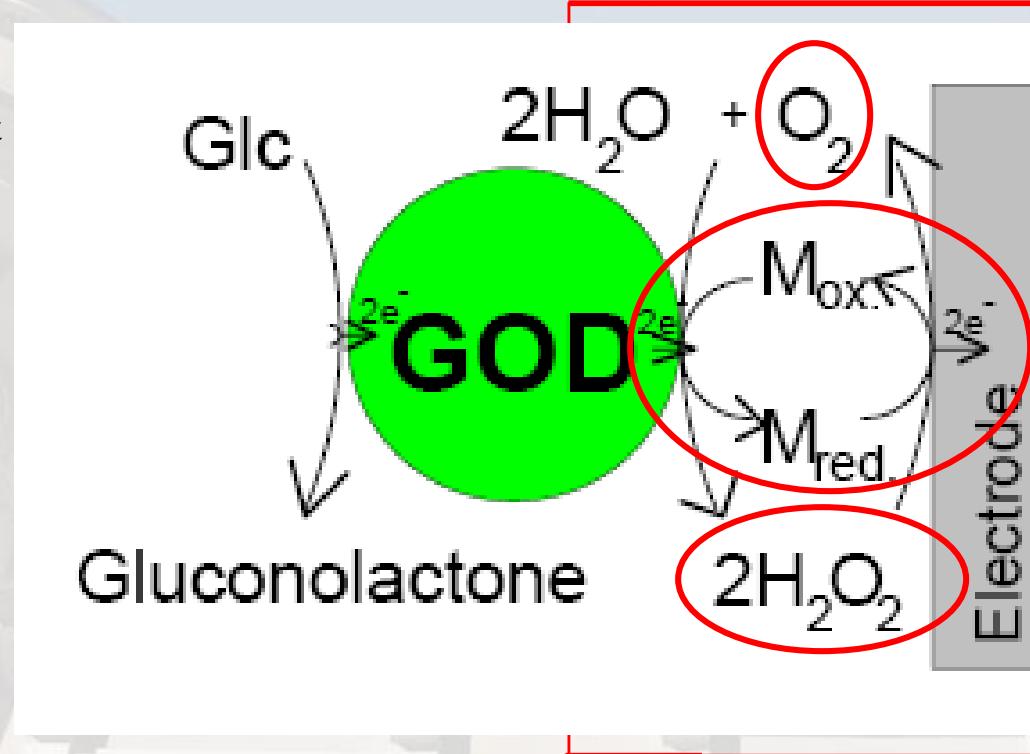
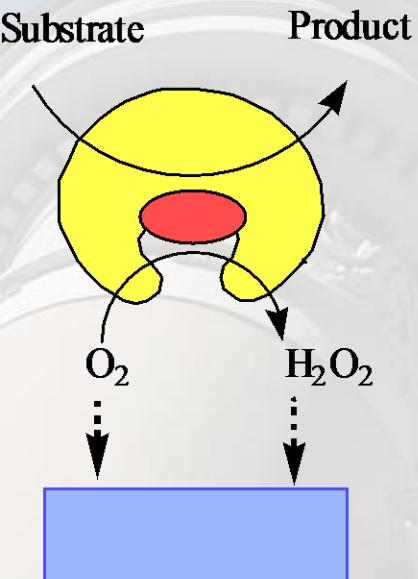


The most promising Enzymes, since has “well established intrinsic electrical circuit” and are able to transfer electrons directly to conducting surfaces.

I=PMS; DCPIP; BQ

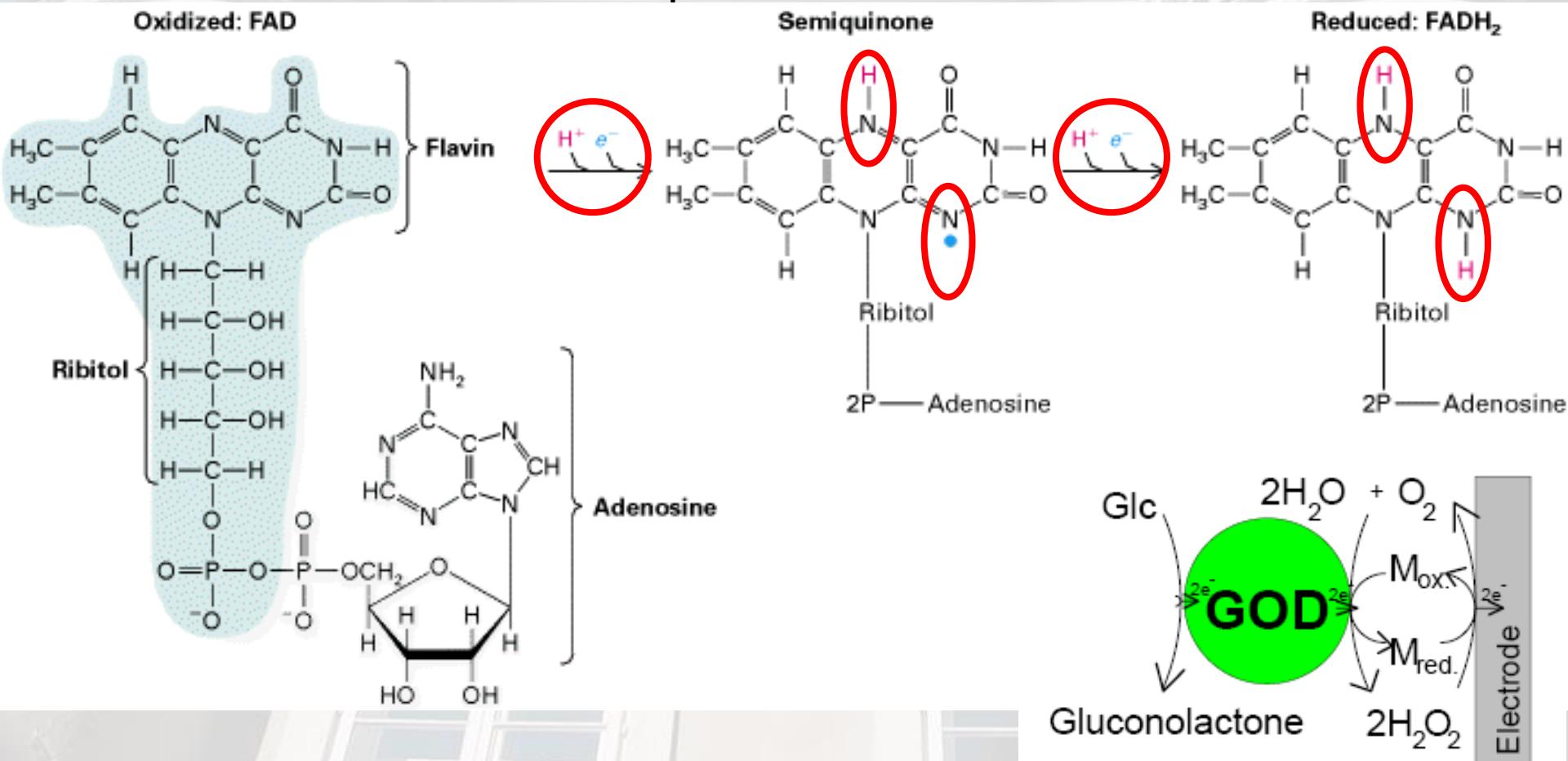


Oxidases – mostly used enzymes in catalytic biosensor design



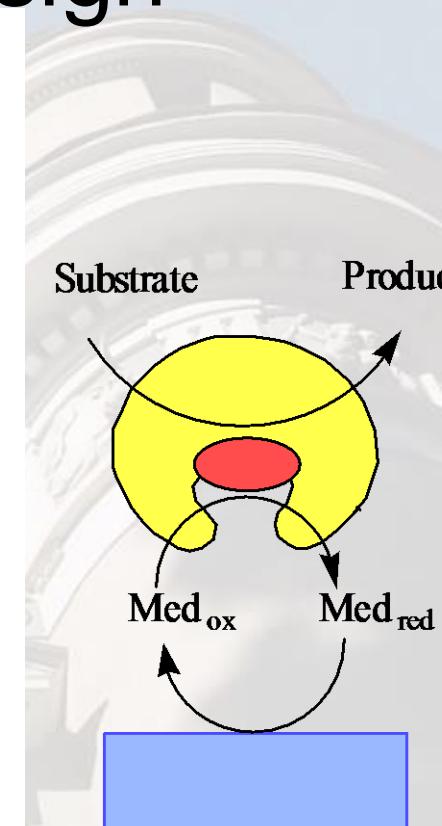
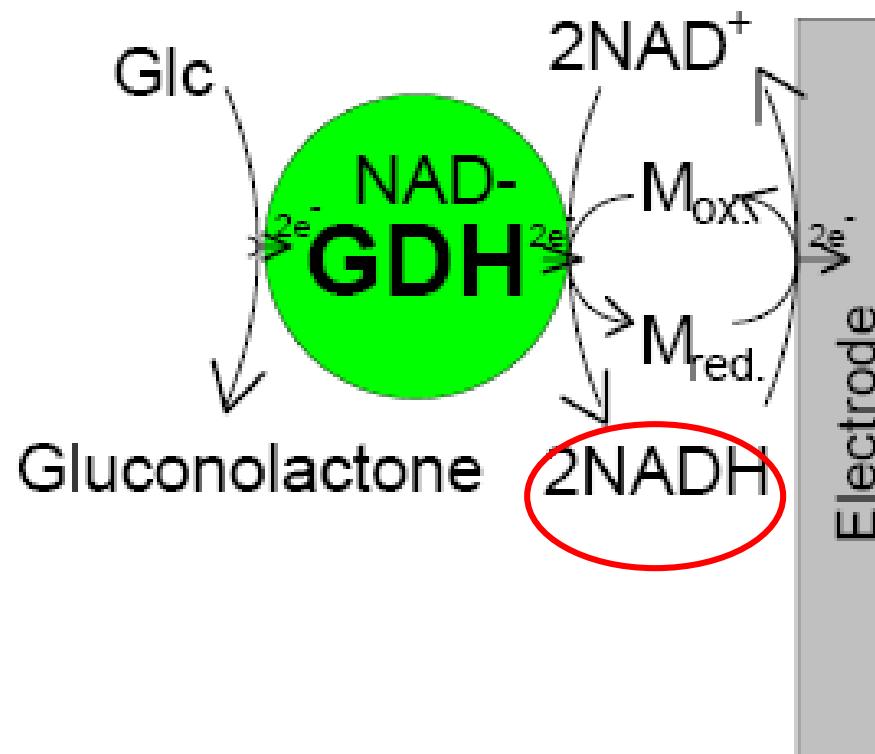
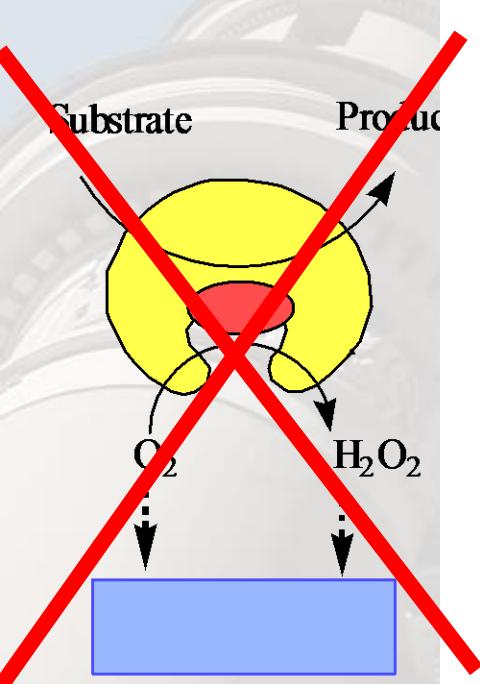
FAD cofactor

- Flavin adenine dinucleotide (FAD) is a redox cofactor involved in several important reactions in metabolism



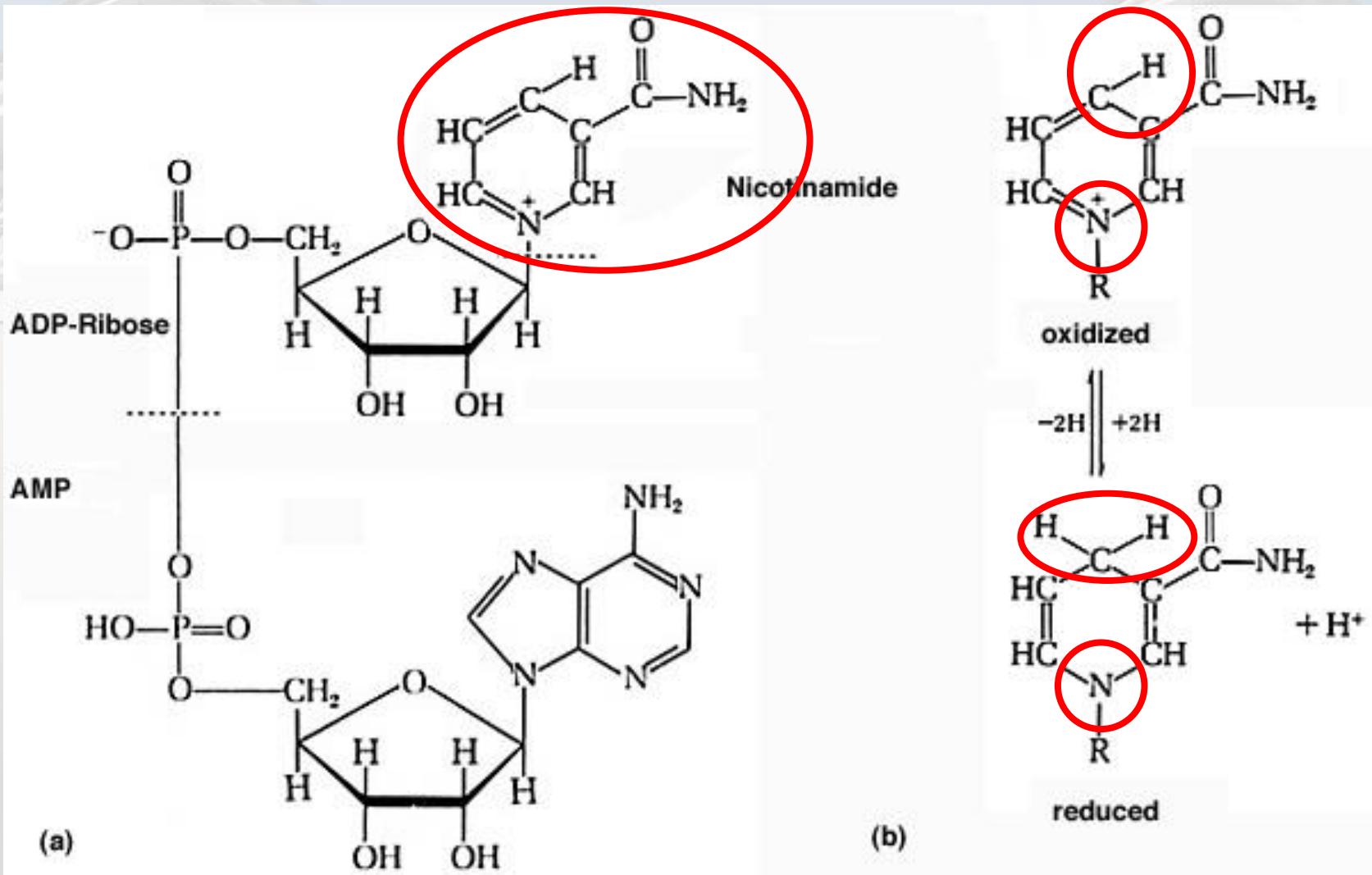


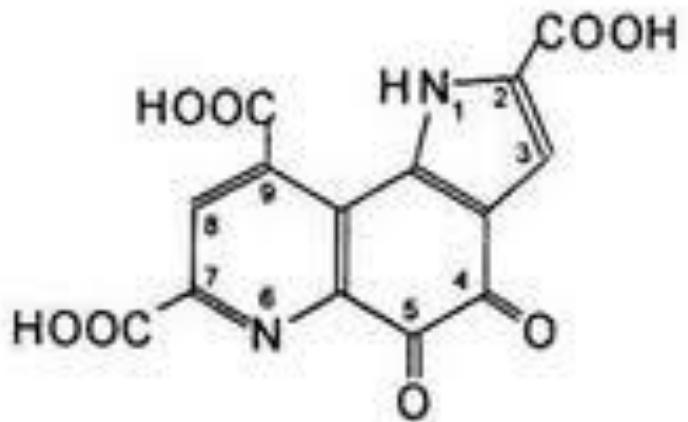
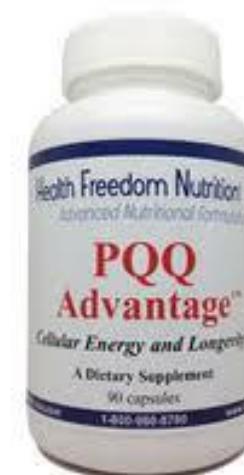
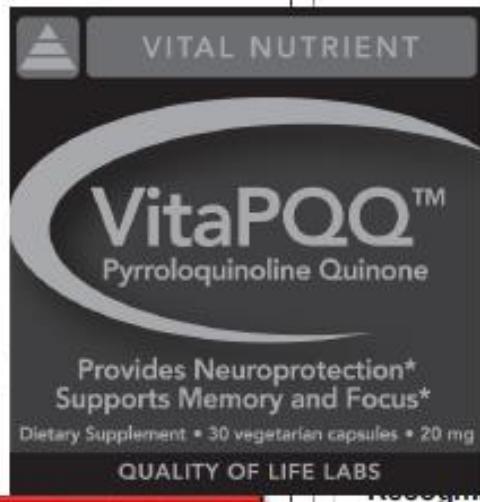
NAD-dependent dehydrogenases in catalytic biosensor design



NAD⁺/NADH cofactor

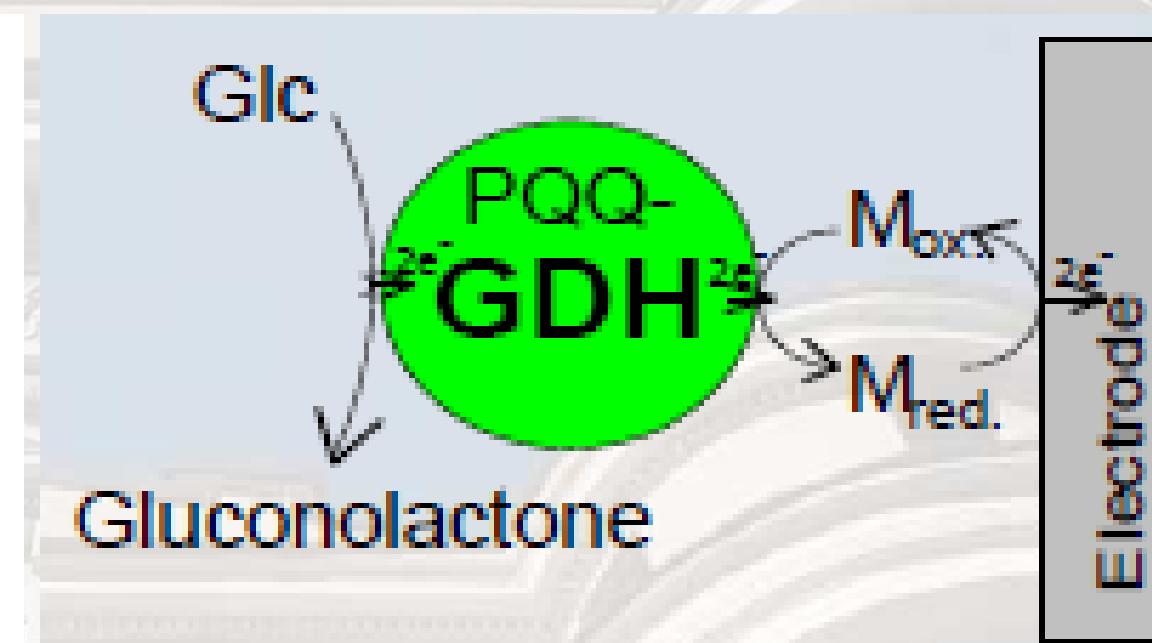
Nicotinamide adenine dinucleotide (NAD)





Pyrroloquinoline quinone [PQQ]

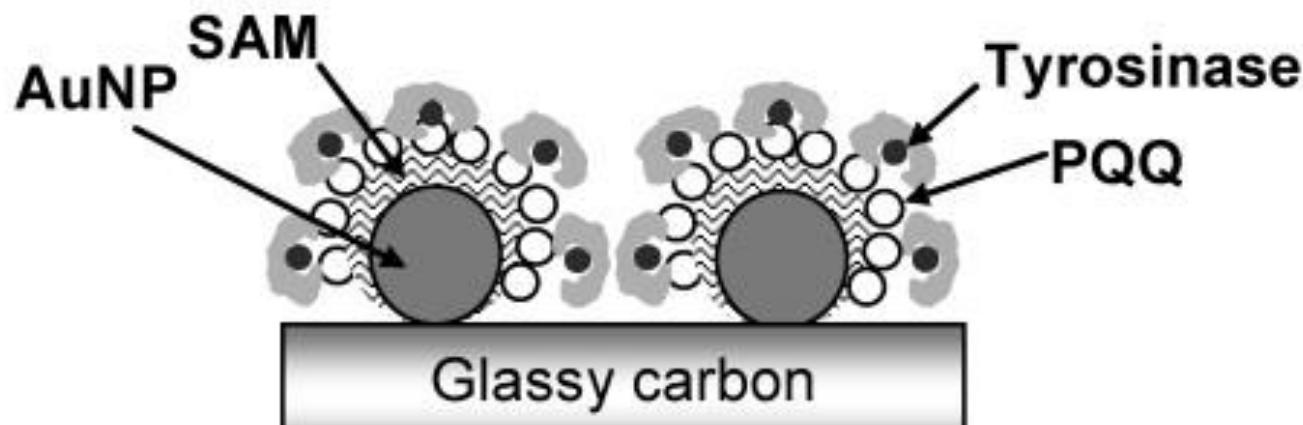
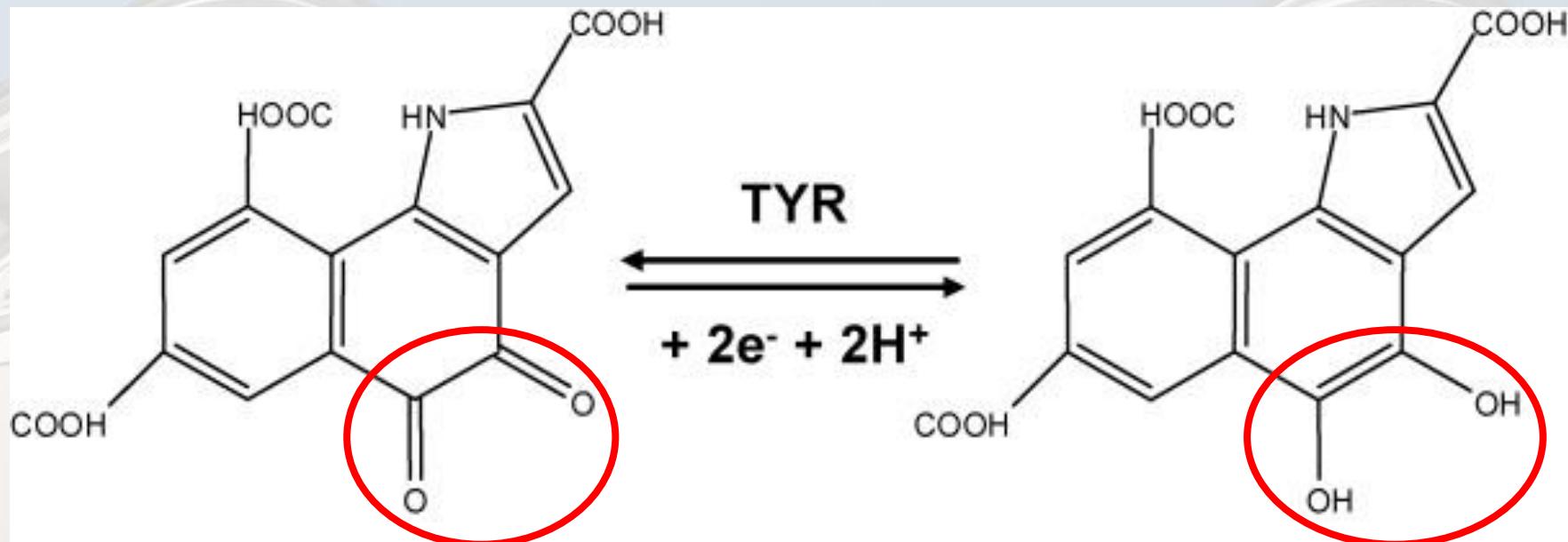
Prosthetic group of bacterial
quinoprotein dehydrogenases





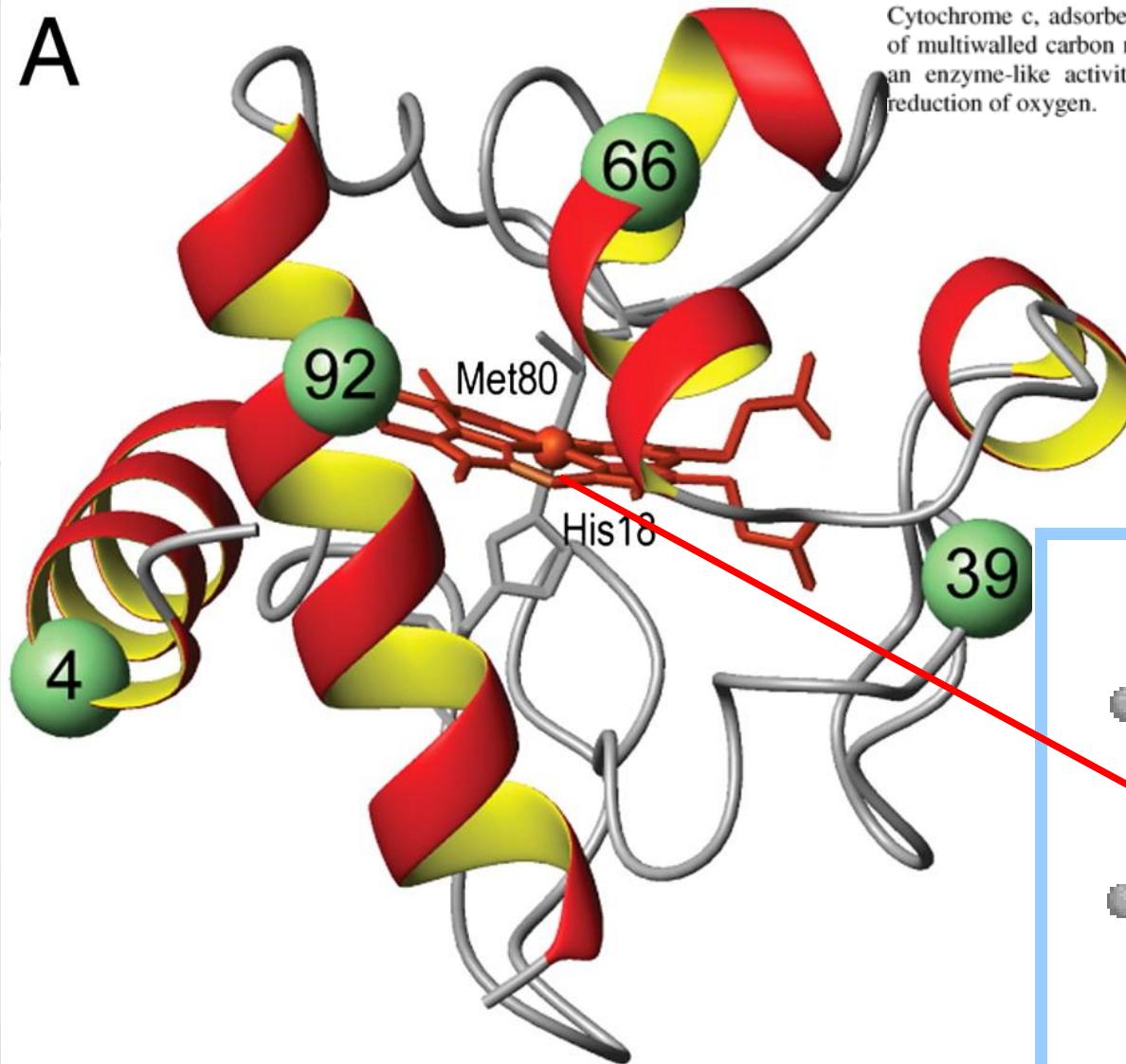
PQQ Cofactor

Pyrroloquinoline quinone (PQQ)

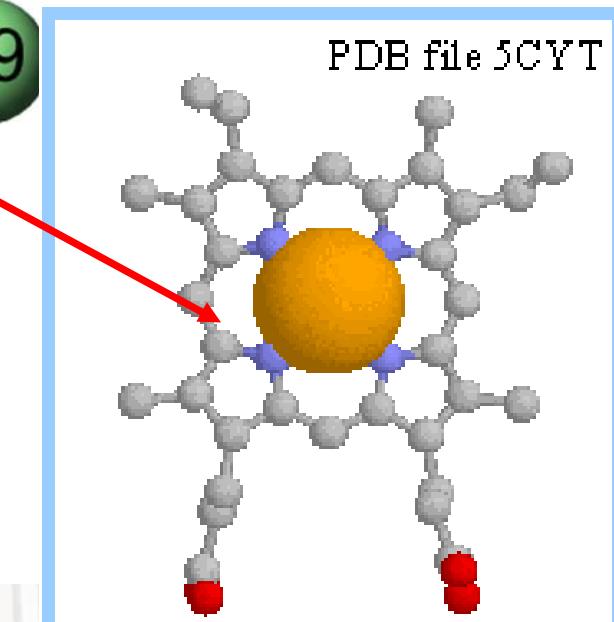
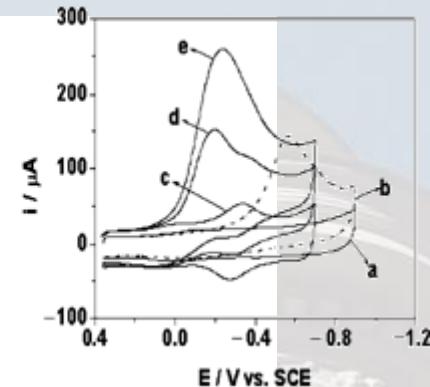


Cytochrome c

A

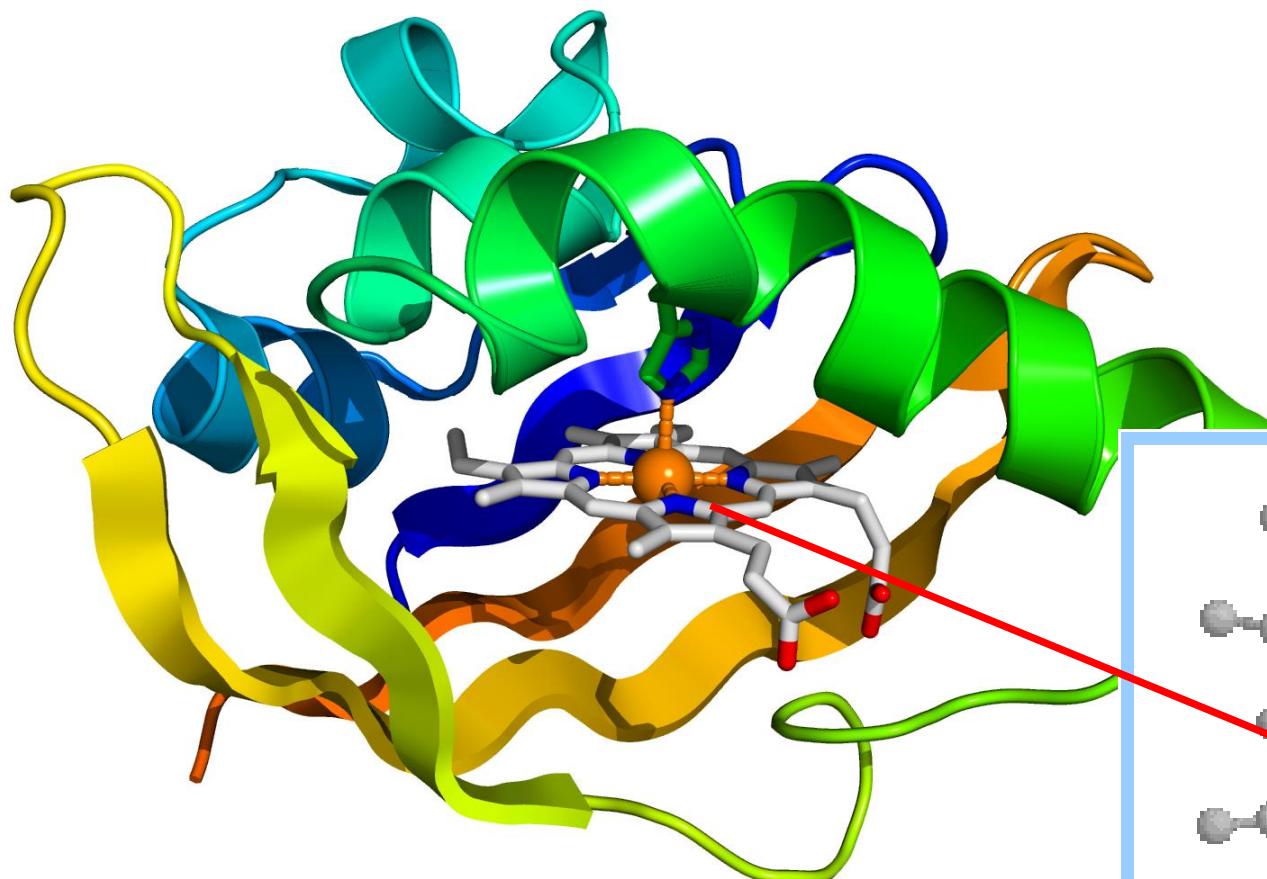


Cytochrome c, adsorbed onto the surface of multiwalled carbon nanotubes, showed an enzyme-like activity to catalyze the reduction of oxygen.

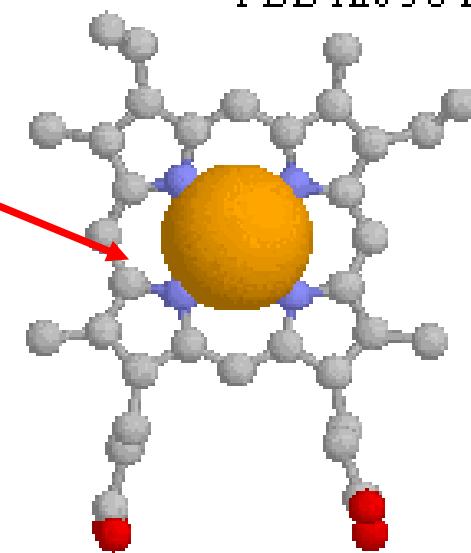




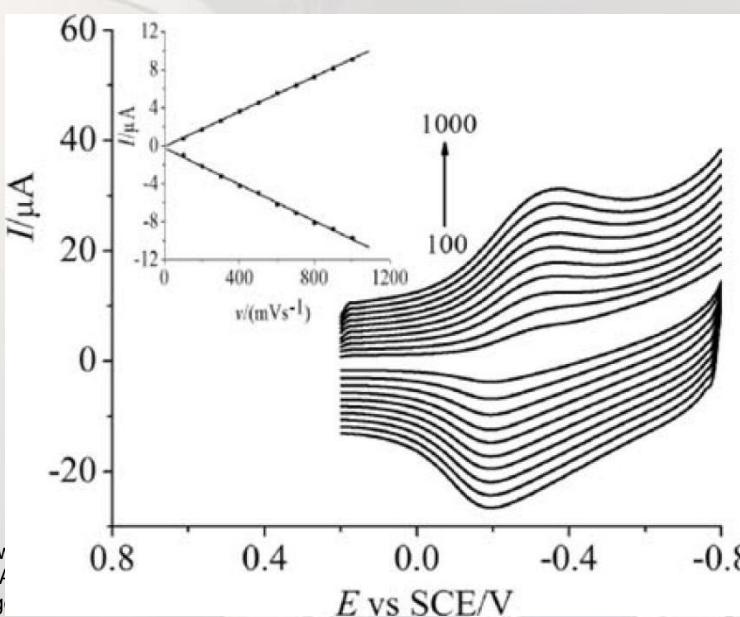
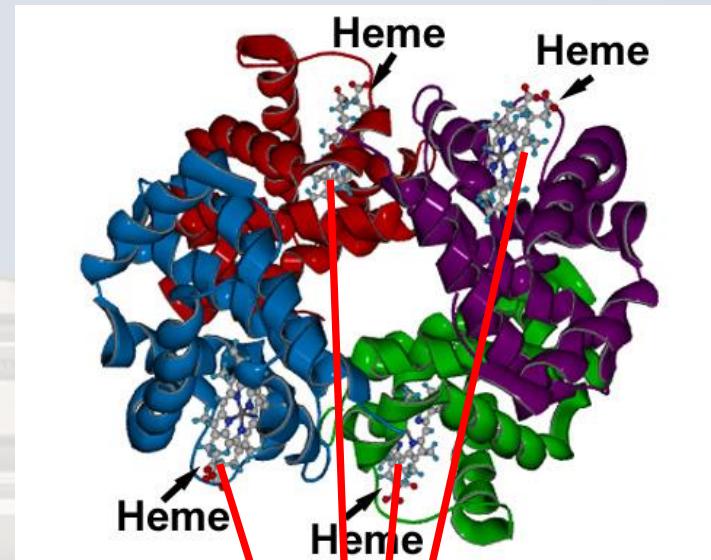
Horseradish peroxidase



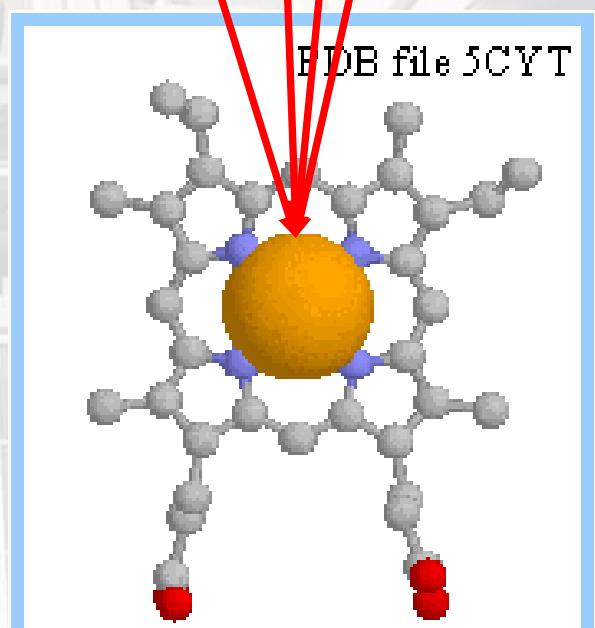
PDB file 5CYT



Hemoglobin



<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC180031452/>



PDB file 5CYT

Heme-c dependent proteins

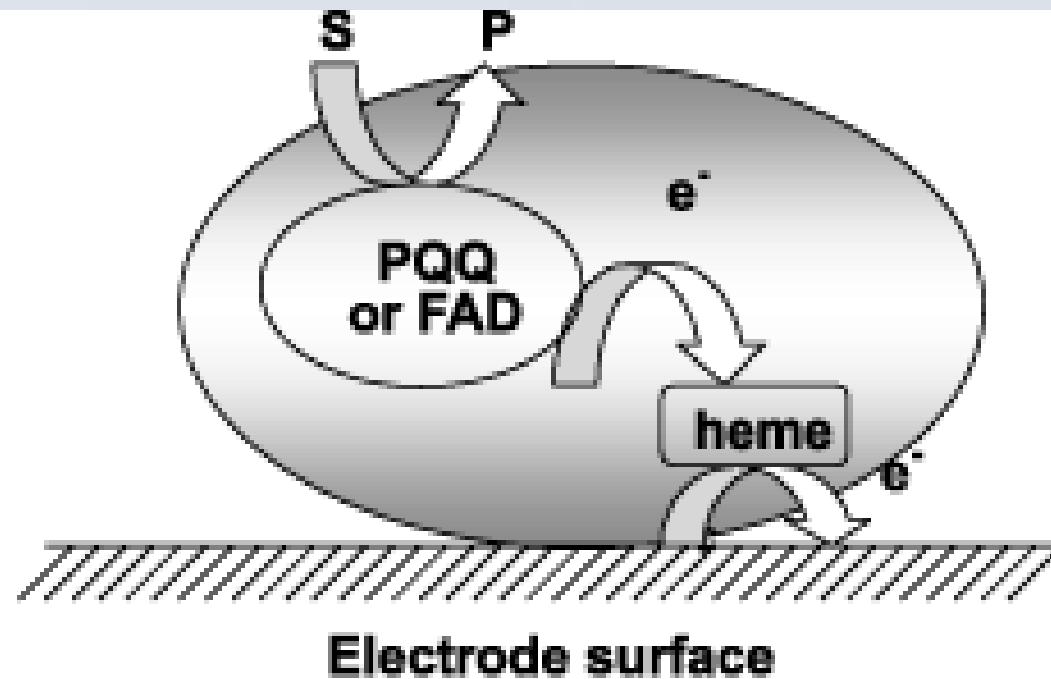
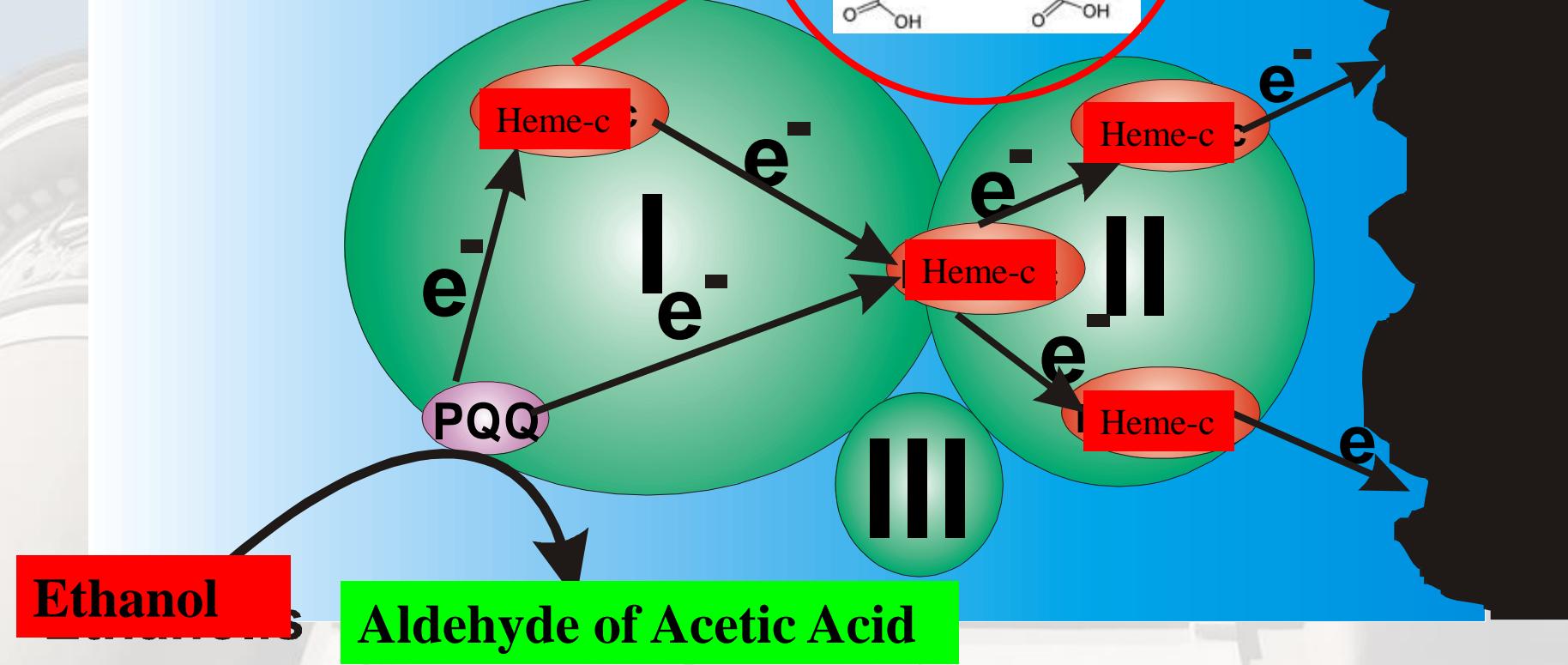
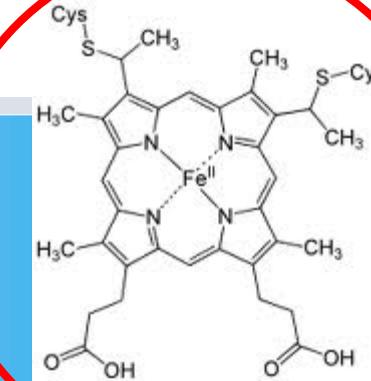


Figure 5. Schematic picture of multi-factor enzymes and the electron transfer pathway.



Ramanavičius A., Habermüller K., Csöregi, E., Laurinavičius V., Schuhmann. W. (1999) Polypyrrole entrapped quinohemoprotein alcohol dehydrogenase. evidence for direct electron transfer via conducting polymer chains, *Analytical Chemistry*, 71, 3581-3586.

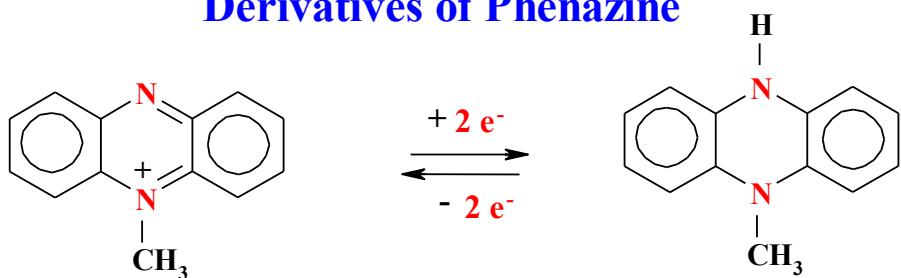
Razumienė J., Niculescu M., Ramanavičius A., Laurinavičius V., Csöregi E. (2002) Direct Bioelectrocatalysis at Carbon Electrodes Modified with Quinohemoprotein Alcohol Dehydrogenase from *Gluconobacter* sp. 33, *Electroanalysis*, 14, 43-49



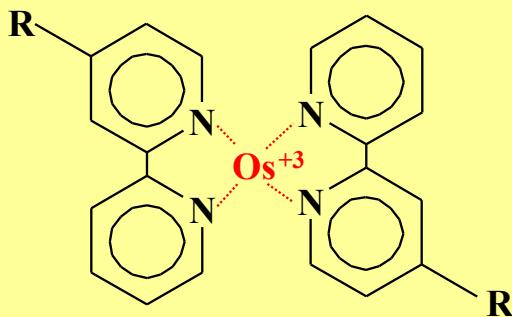


Some Red-Ox Mediators

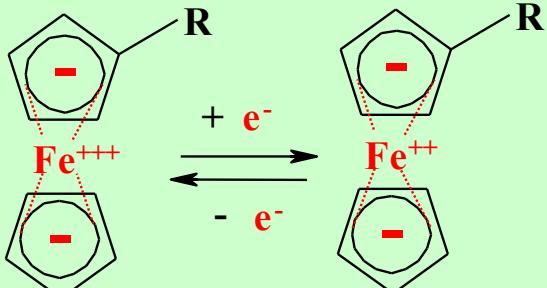
Derivatives of Phenazine



Derivatives of Os-bipyridine complex



Derivatives of Ferrocene

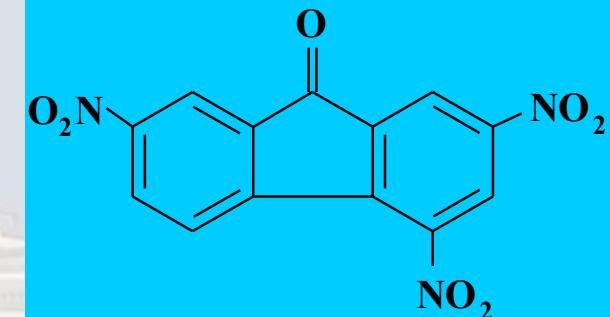


Prussian Blue

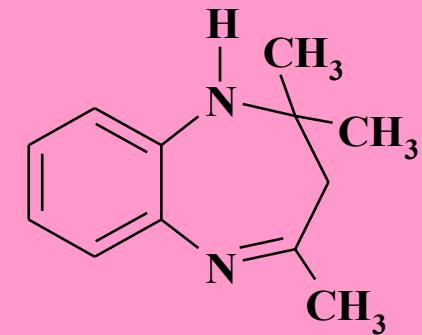


some RedOx Medi

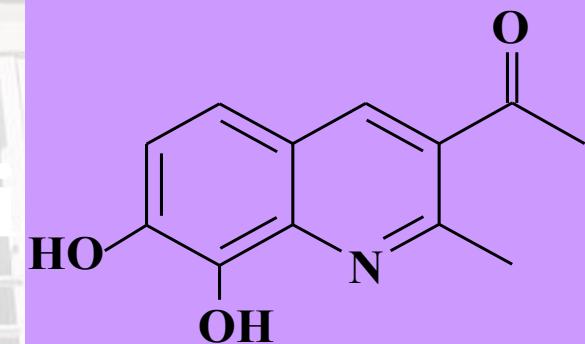
Fluorenone



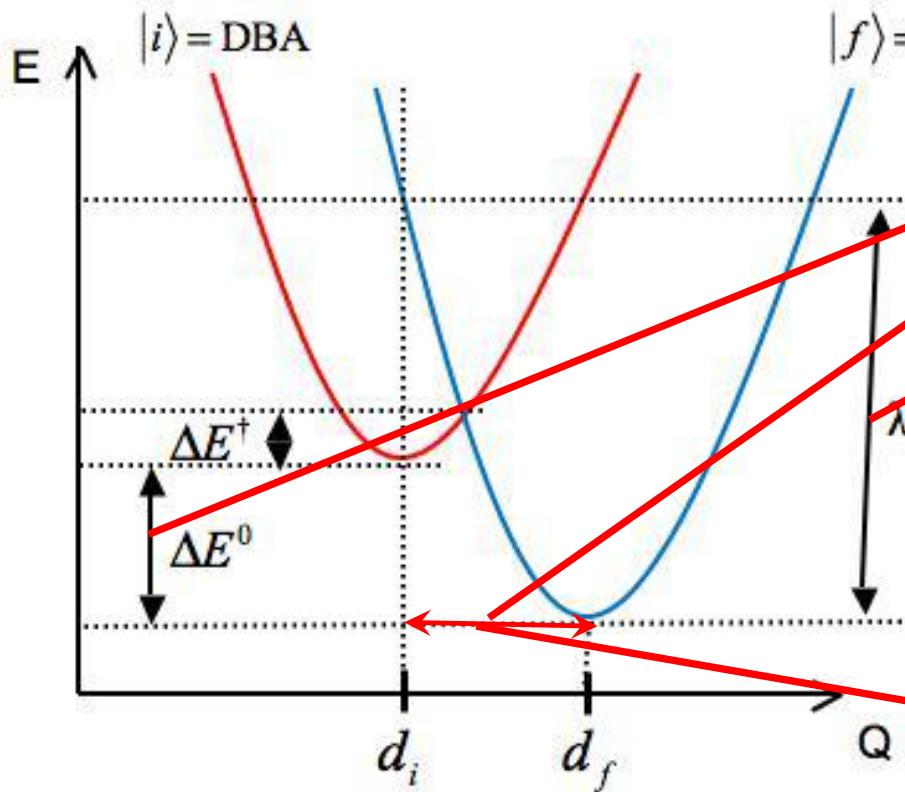
Derivatives of benzodiazepine



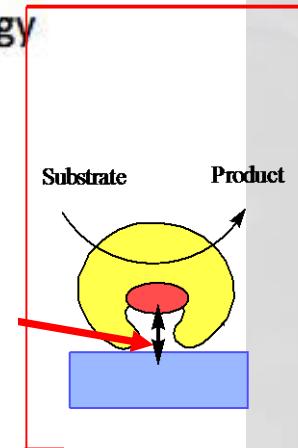
Derivatives of quinones



Electron transfer of efficiency // Marcus-Hush theory



thermodynamic driving force
 $k_s \propto e^{-\beta(d-d_0)} e^{\frac{-(\Delta G^\circ + \lambda)^2}{4RT\lambda}}$
 distance
 λ : reorganization energy
 ΔE^0 : energy of reaction or driving force
 ΔE^\dagger : activation energy

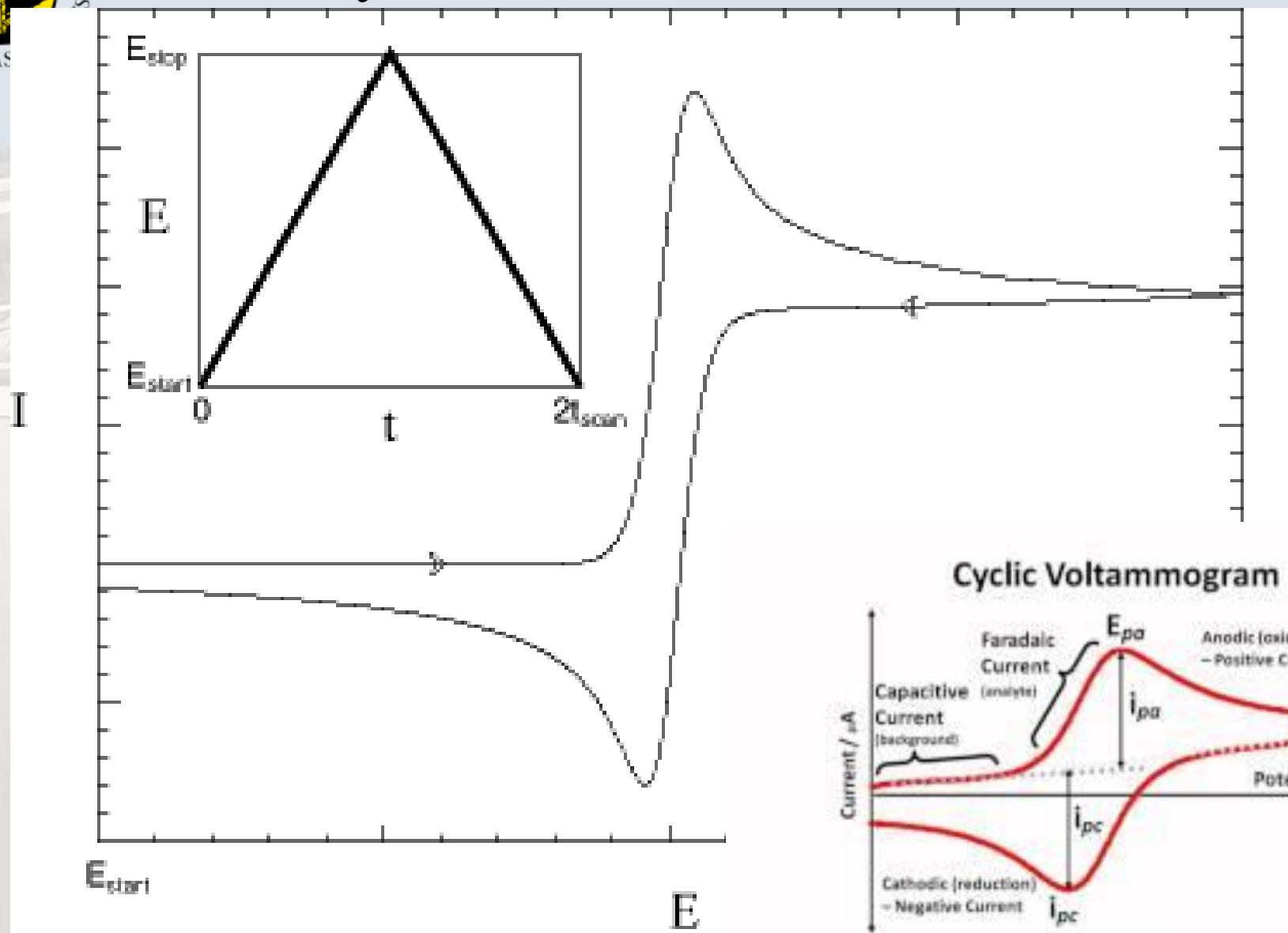


Electron transfer (ET) reactions involve the movement of an electron from one molecular species (the donor) to another (the acceptor)

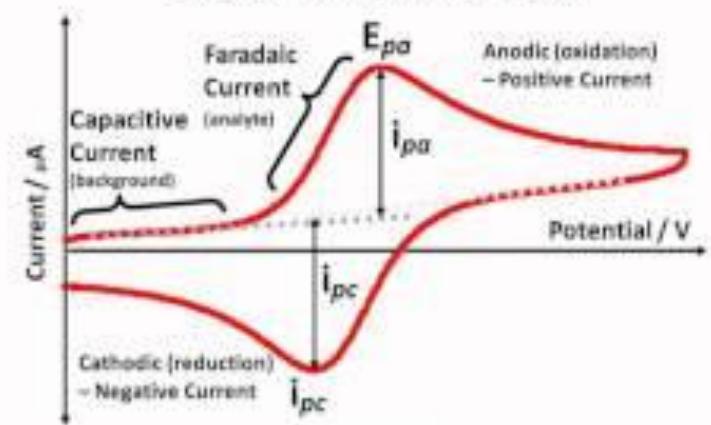
$$k_{ET} = |V_{el}|^2 \sqrt{\frac{\pi}{h^2 k_B T \lambda}} \exp \left\{ -\frac{(\Delta E^0 - \lambda)}{4\lambda k_B T} \right\}$$



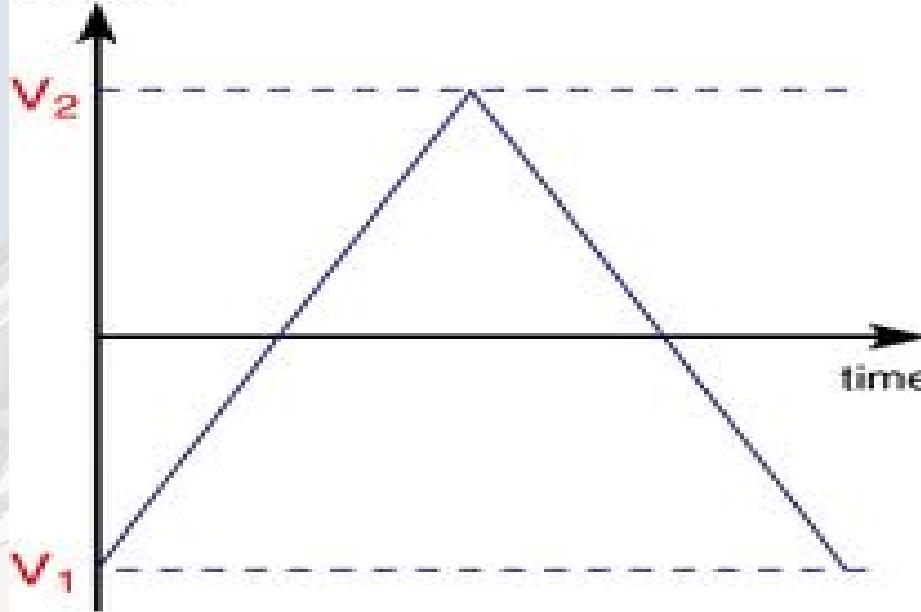
Electron transfer of efficiency // Potentiodynamic methods



Cyclic Voltammogram



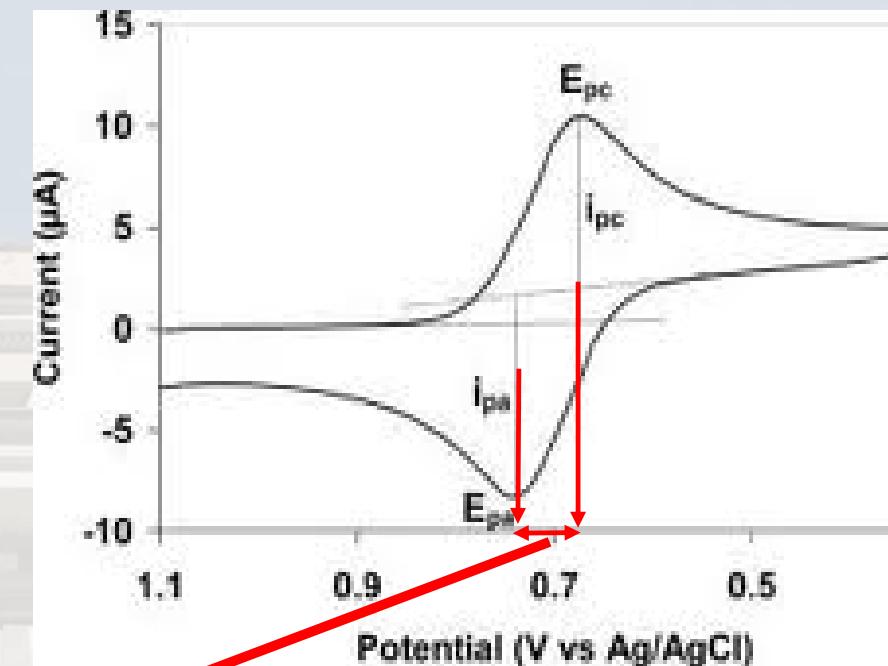
Electron transfer of efficiency // Cyclic voltammetry



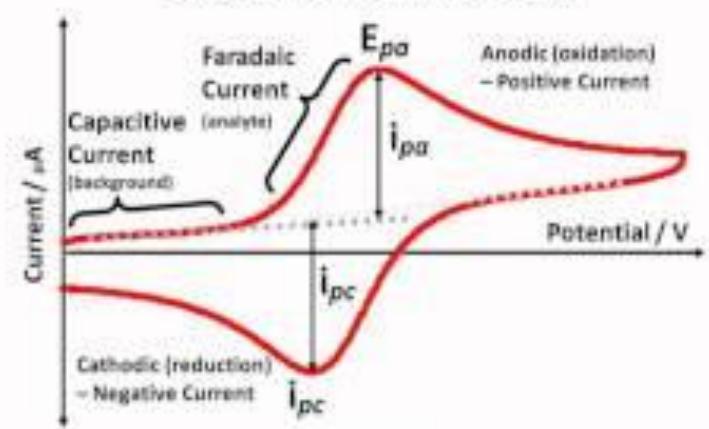
Nicholson, Kochi's method

$$k^o = 2.18 \left[\alpha v n F D_o / RT \right]^{1/2} \exp \left[-\frac{\alpha^2 n F \Delta E_p}{RT} \right]$$

Heterogeneous electron transfer constant k^o

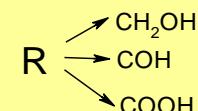
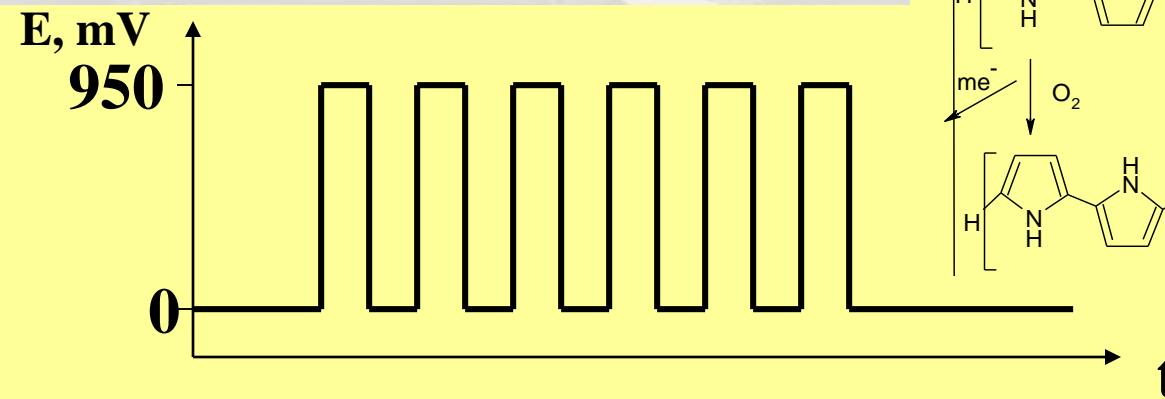
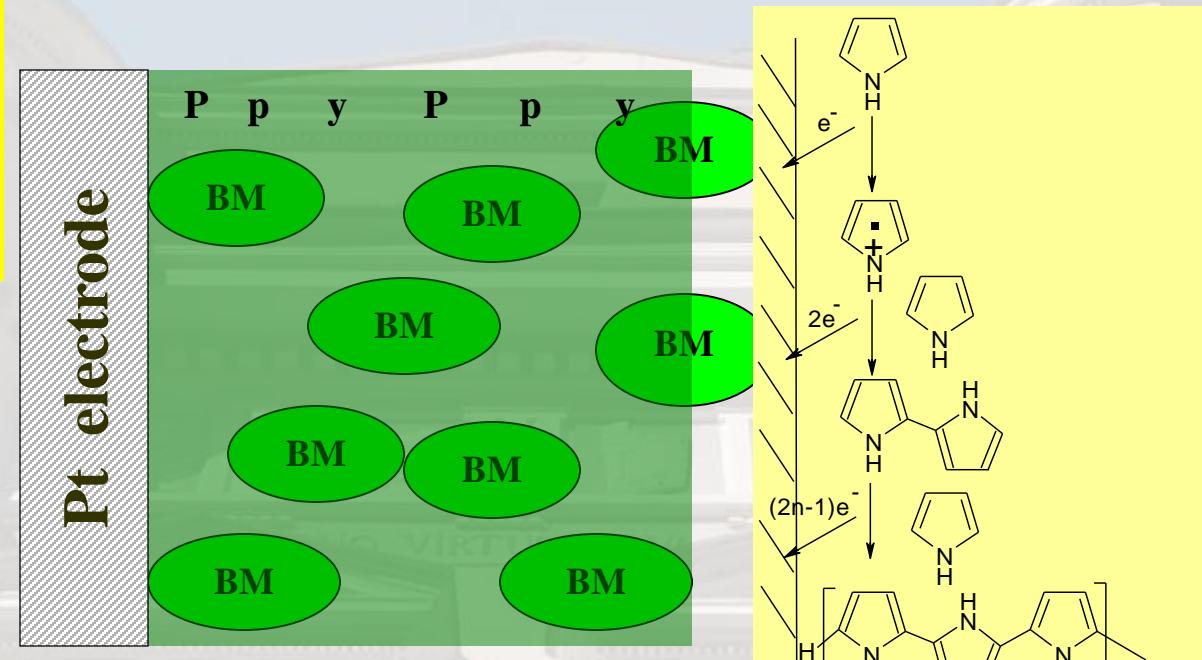


Cyclic Voltammogram



Immobilization of bio-molecules within electrochemically formed polypyrrole (Entrapment of bio-molecules within Ppy)

A. Ramanavicius, Y. Oztekin, A. Ramanaviciene
 Electrochemical Formation of Polypyrrole-based Layer for Immunosensor Design.
Sensors and Actuators B-Chemical 2014, 197, 237–243.

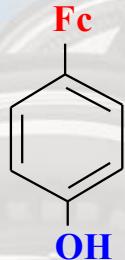




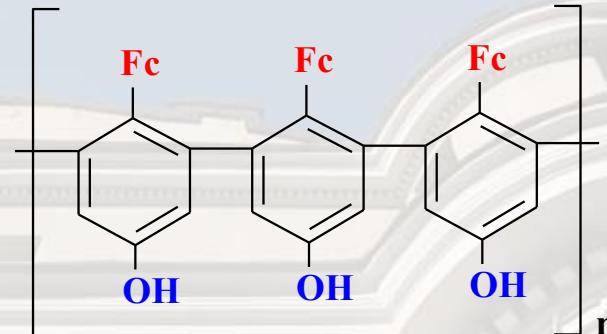
Electrochemically polymerisable Compounds with Red-Ox Mediators

Ferrocenepheno^l

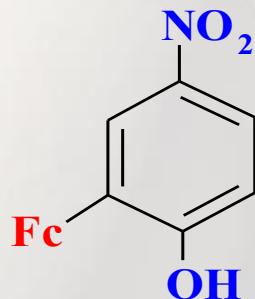
(FP)



Electropolymerisation

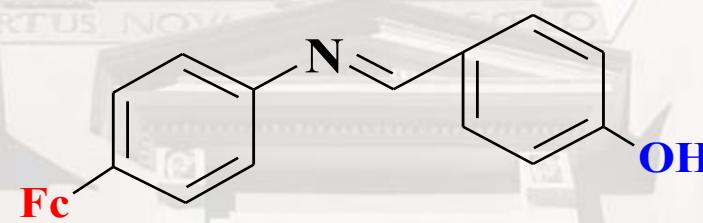


2-Ferrocenyl-4-nitropheno^l



(FcNO₂)

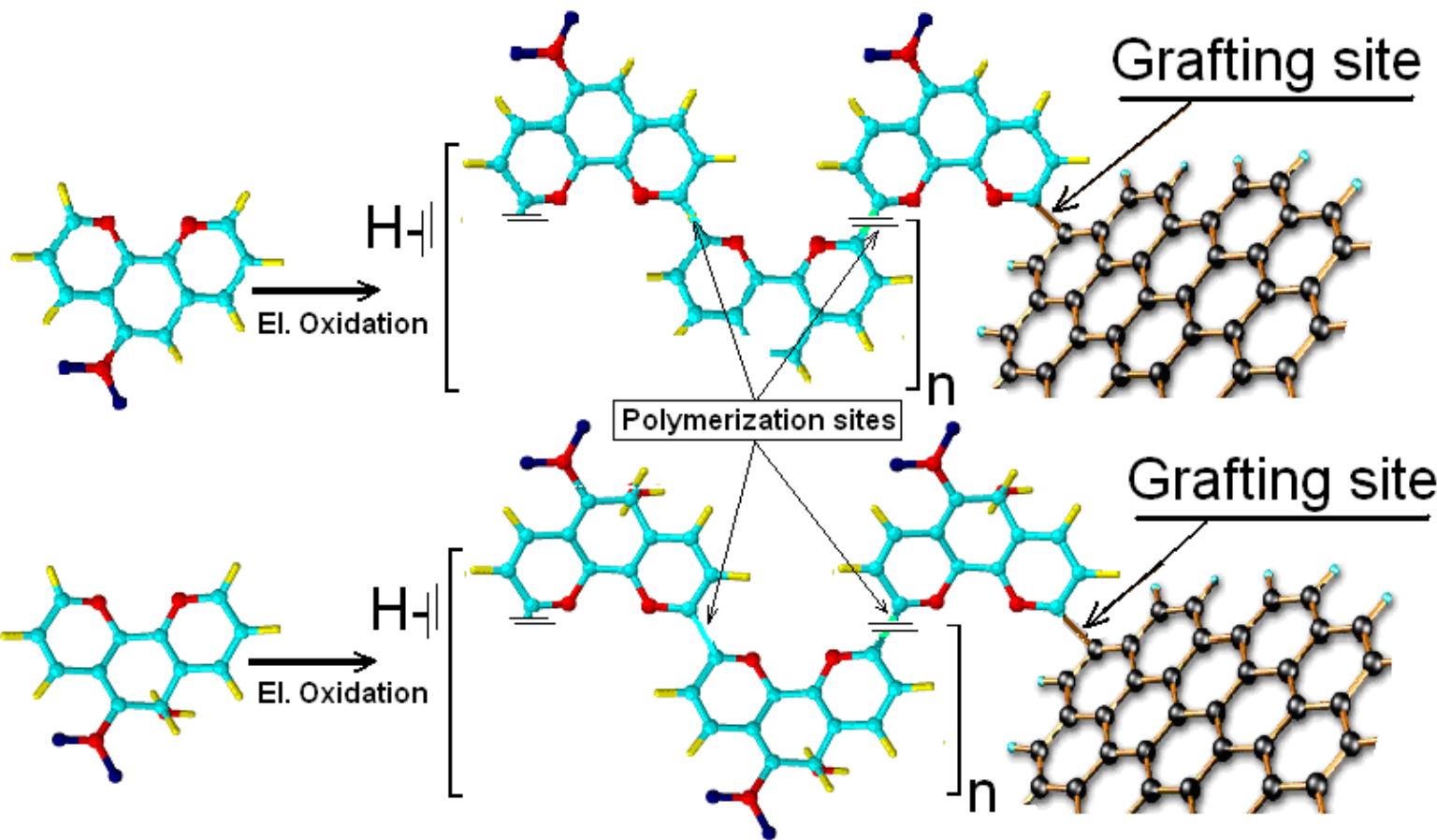
N-(4-Hydroxybenzylidene)-4-ferrocenylaniline

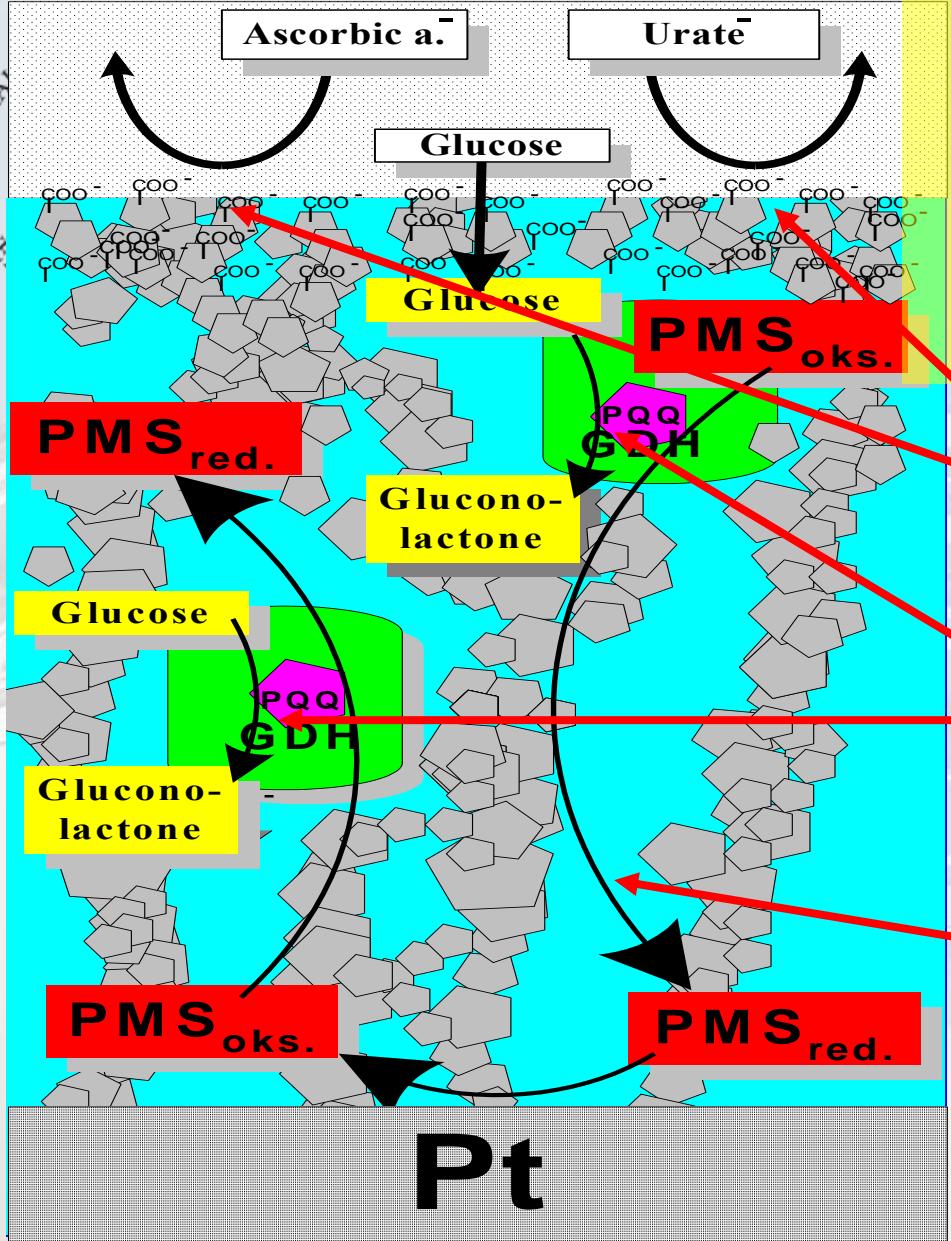


(FP1)

Electrochemical
polymerisation

“Grafted” conducting polymers



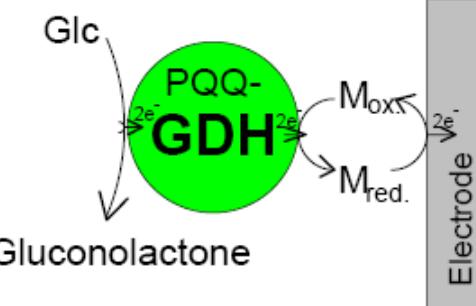


PMS mediated biosensor. Based on PQQ-GDH entrapped within conducting polymer

Protection from interfering chemicals, by over-oxidized Ppy layer

Catalytic conversion of analyte

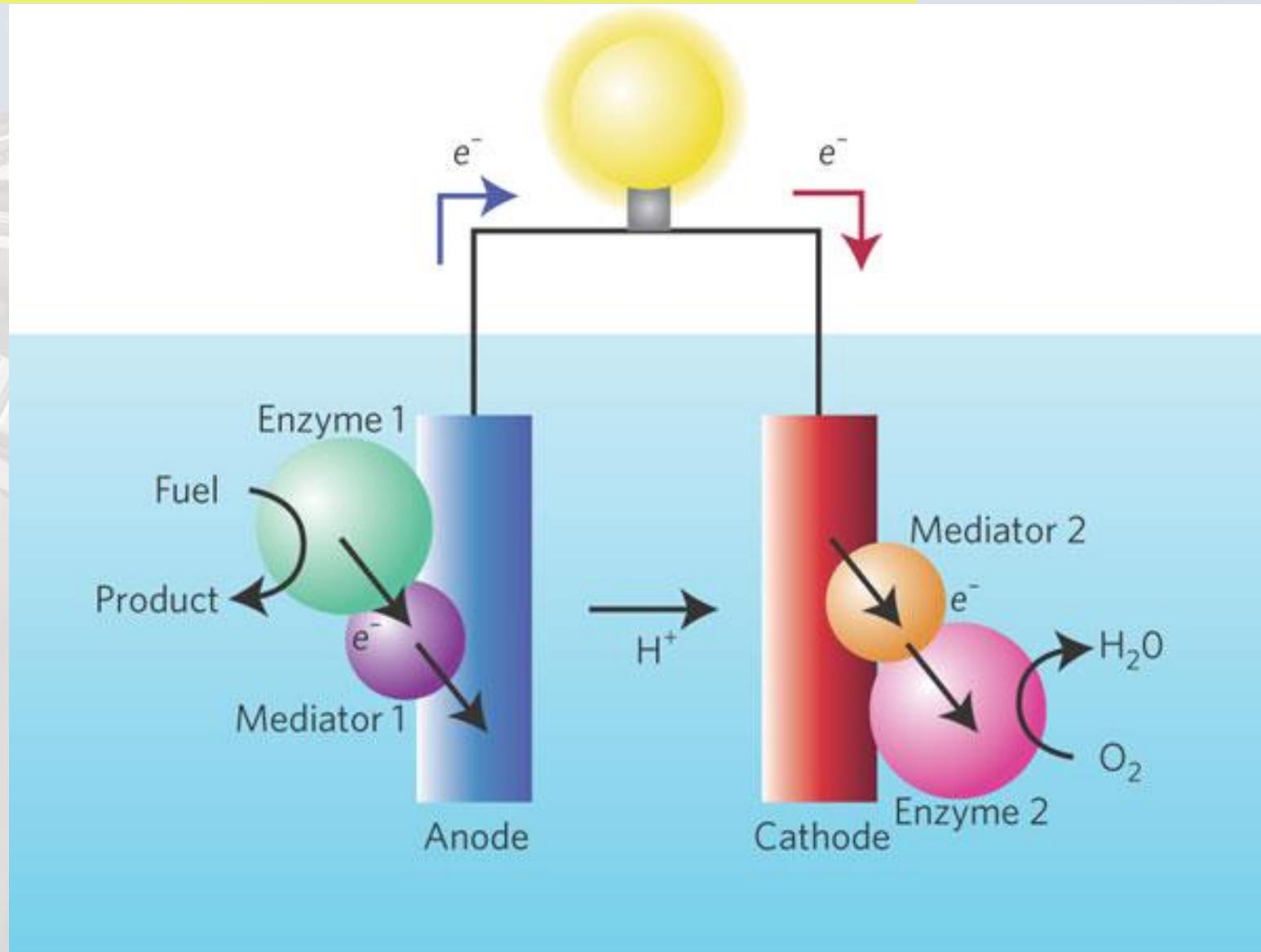
Transfer of electrons from enzyme to conducting surface by diffusing “electron shuttles”



Ramanavicius A. (2000) Electrochemical study of permeability and charge-transfer in polypyrrole films, *Biologija*, 2, 64-66.

Laurinavicius V., Razumiene J., Ramanavicius A., Ryabov A.D., (2004) Wiring of PQQ-dehydrogenases, *Biosensors & Bioelectronics* 20 (6), 1217-1222.

Enzymatic Biofuel cells



Advantages of biofuel cells (1)



(Bio)fuel cell

Really green
energy

Most usual generators of electricity

Chemical or nuclear energy

Thermal energy -

Mechanical energy

Electrical energy





Potential range

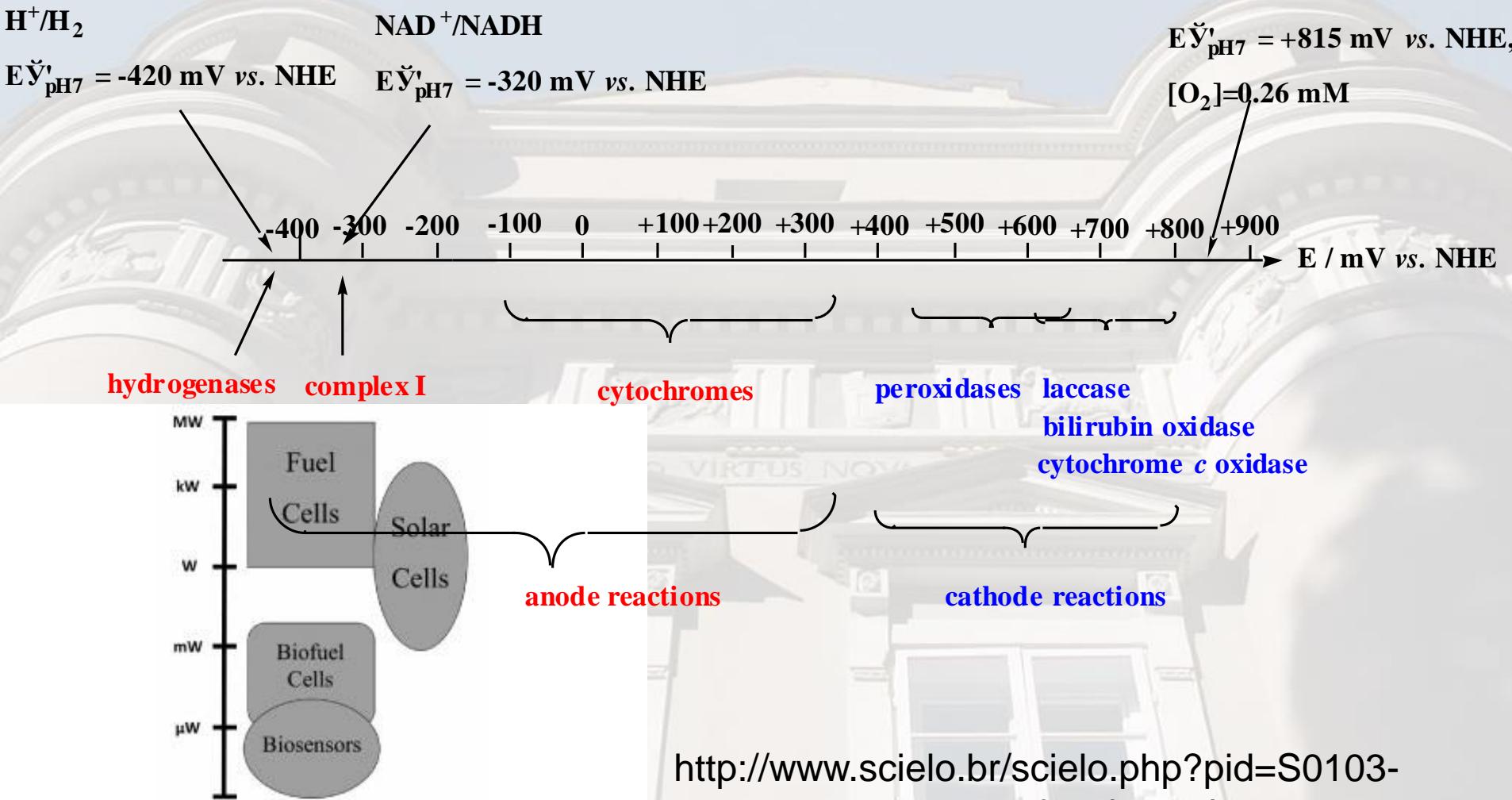
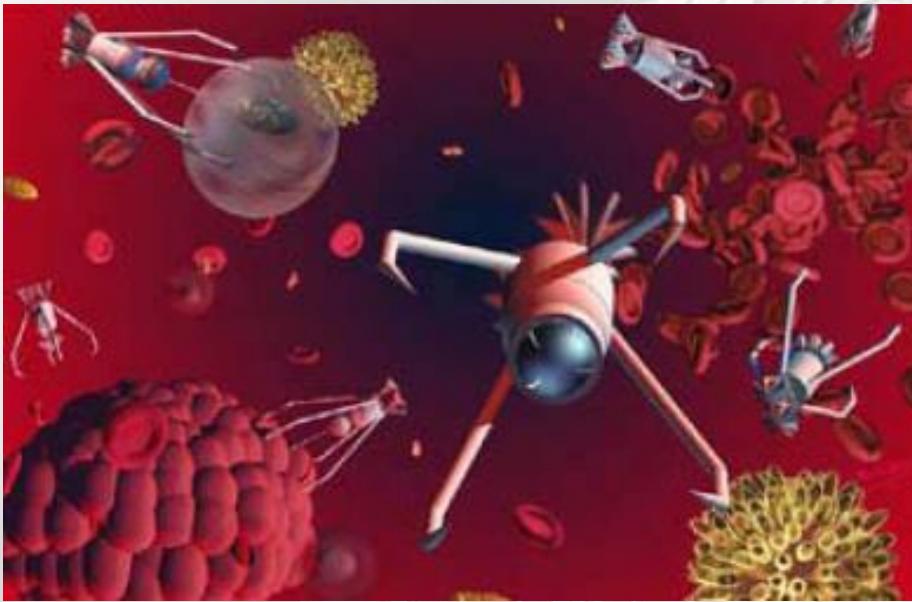
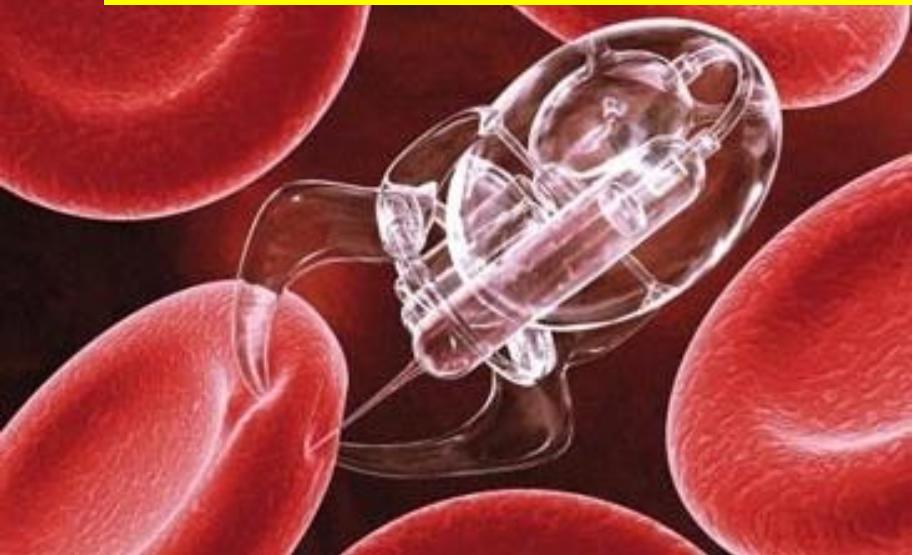


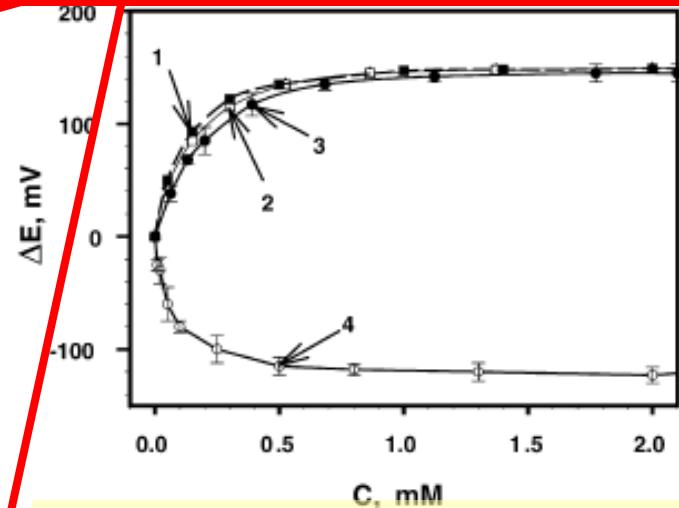
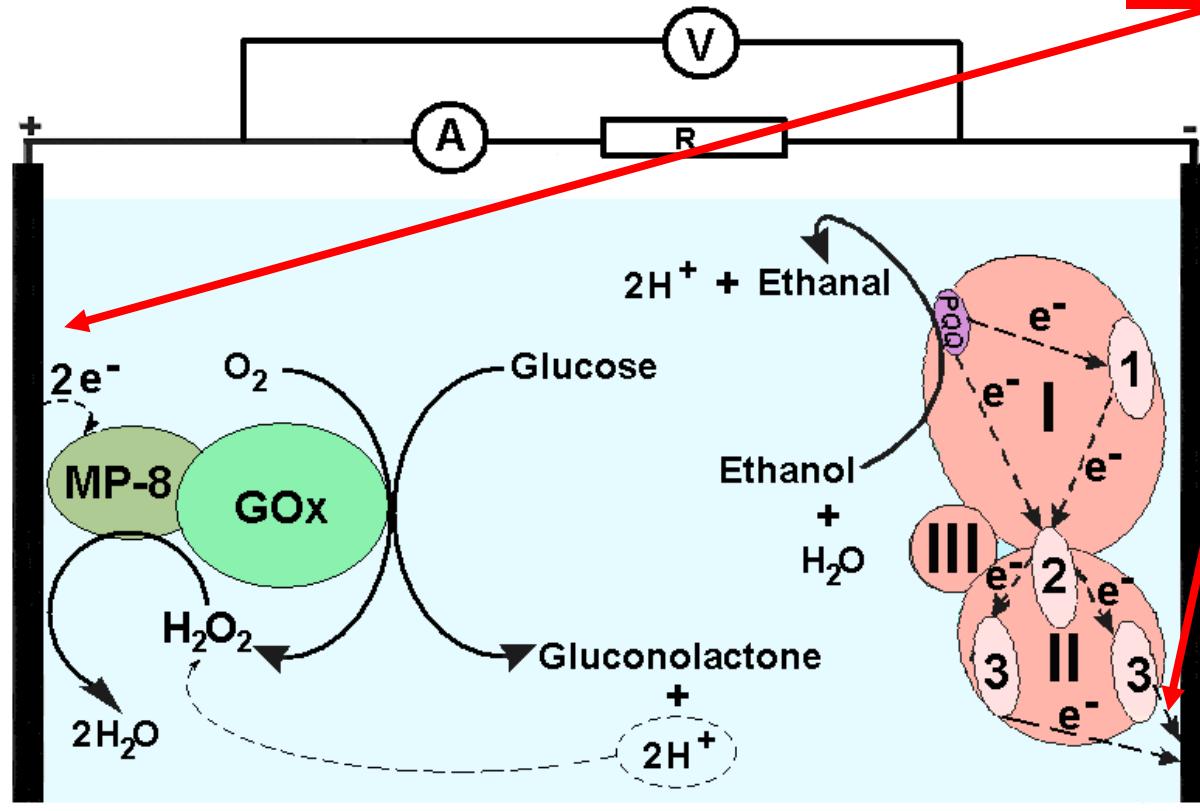
Figure 3. Schematic representation of the power range that some of the alternative energy production methods provide (adapted from reference 8)

NanoRobots, how to power them?



Enzymatic Biofuel cells

Direct Electron transfer



Potentials of (1) MP-8 functionalized cathode as function of hydrogen peroxide concentration; (2) MP-8/GOx functionalized carbon rod electrode as a function of hydrogen peroxide concentration; (3) MP-8/GOx functionalized carbon rod electrode as a function of glucose concentration; (4) QH-ADH functionalized anode as a function of ethanol concentration. Investigations performed in 50 mM Na acetate solution, pH 6,

Ramanavicius A., Kausaite A., Ramanaviciene A., (2005) Biofuel cell based on direct bioelectrocatalysis, *Biosensors and Bioelectronics*, 20: 1962-1967.

Ramanavicius A., Kausaite A., Ramanaviciene A., (2006) Potentiometric Study of Quinohemoprotein Alcohol Dehydrogenase Immobilized on the Carbon Rod Electrode, *Sensors and Actuators B: Chemical*, 113, 435-444.



Thank YOU for attention!

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e-mail: Arunas.Ramanavicius@chf.vu.lt.*

Main activities: Development of bio-, immuno- and DNA-sensors; Development of bio-analytical methods for the food, environmental analysis and biomedical application; Synthesis and application of conducting polymers.